



Research Journal of Pharmaceutical, Biological and Chemical Sciences

Treatment of Aqueous Solution of Phenol by Adsorption on Sodium Zeolite

Saravanakumar K¹ and Kumar A^{2*}

¹Dept.of Chemical Engineering, St.Peter's Engineering College and Research Institute, Chennai-54, India.

²C.Abdul Hakeem College of Engineering & Technology, Melvisharam-632 509, Vellore-Dist, India.

ABSTRACT

The adsorption of phenol by sodium zeolite was investigated to assess its possible use as an adsorbent. The adsorption of phenol is found to be the most important problem being faced by many industries like textiles, pharmaceuticals, petrochemical industries etc. The optimum conditions for maximum adsorption in terms of pH, contact time, amount of the adsorbent and adsorbent equilibrium were identified. The Freundlich and Langmuir models were used for the mathematical description of adsorption equilibrium and it was found that the experimental data fitted very well to the Langmuir model. The results indicate that sodium zeolite can be employed as a low cost alternative to commercial adsorbent in the removal of phenol from aqueous solutions.

Key words: phenol, adsorption, sodium zeolite, Freundlich, Langmuir.

***Corresponding author**

E-mail: ayanakumar@yahoo.com.



INTRODUCTION

The waste solution generated from coal conversion processes contains substantial amounts of phenol, i.e. 200-600 mg/l [20], which is discharged into natural water streams, contaminating the ground water. Phenol is considered to have toxic effects on human health even when present in small concentrations [11]. The ingestion of such contaminated water in the human body causes protein degeneration, tissue erosion, and paralysis of the central nervous system and also damages the kidney, liver and pancreas [11]. The threshold value of phenol in water is 4000 µg/l.

Since 1860, phenol has been in production, with its basic use as an antiseptic. During late 19th century and thereafter the use of phenol has been further extended to the synthesis of dyes, aspirin, plastics, pharmaceuticals, and petrochemical and pesticide chemical industries. In fact, by 2001, the global phenol production has reached an impressive 7.8 million tons [17]. Normally, discharges from the aforesaid phenol-yielding industries find their way to water bodies and subsequently affect the water quality adversely. As per the studies by various authors, phenol and its derivatives have been reported to cause undesirable and deleterious effect in water, even at a concentration as low as 0.1 ppm [14]. In fact, the adverse effects of phenol have been observed on the central nervous system, cardiovascular system as well as urino-genital systems of human being, often expressed by the multiple symptoms: convulsions, coma, cardiac disorders, respiratory failure and collapse [1]. As per literature, the various analytic methods attempted for dephenolation of wastewater include steam stripping, solvent extraction, oxidation (O_3 , H_2O_2 , and ClO_2), ion exchange, biodegradation and adsorption methods [13, 21, 22]. Out of all these, adsorption methods are the most-widely used techniques and activated carbon has been the predominant adsorbent all over the world [16, 8]. However, due to its high cost and low regeneration capacity, since the last three decades, extensive researches have been directed towards investigating the adsorption characteristics and potentials of cheaper materials and solid wastes, such as fly ash, peat, soil, rice husk, sawdust, bagasse and so forth [18, 23, 4, 10].

Human consumption of phenol contaminant water can cause severe pain leading to damage of the capillaries ultimately causing death. Their presence in water supplies is noticed as bad taste and odor [16]. Therefore, Environmental Protection Agency regulations call for lowering phenol content in wastewater to less than 1 mg/L [7]. There are many methods, such as, adsorption, microbial degradation, chemical oxidation, precipitation, ion exchange and solvent extraction to remove phenol from aqueous solution. Adsorption is an effective separation process for treating industrial and domestic effluents.

In another experiment, it was reported that the adsorption of phenol onto chitin was endothermic and increased with temperature [6]. The increase in active bentonites mass or initial pH of the solution also resulted in more phenol removal from solution [3]. Other adsorbents used to remove phenol from aqueous solution are bentonite [4], silica gel, activated alumina and activated carbon [19], titanium oxide (TiO_2) [5].

The objective of this study was to produce water hyacinth ash as an adsorbent for adsorption of phenol, to characterize it and to investigate the effects of initial 17 concentrations, contact time, pH and adsorbent dosage on the adsorption of phenol on water hyacinth ash. Equilibrium isotherm data were fitted to Langmuir and Freundlich equations and constants of isotherm equations were determined. Adsorption kinetics of phenol onto water hyacinth ash was also analyzed by using pseudo-first-order and pseudo-second-order models to data.

Therefore, it is considered necessary to remove the phenol from industrial effluent before discharging into the water stream. Though bioaccumulation and biosorption are mainly preferred for this task as the phenols are easily biodegradable, adsorption onto sodium zeolite is also effective. In this study, sodium zeolite is used as an adsorbent to remove phenol from aqueous solution. Sodium zeolite is available in all the commercial chemical agents and was found to be cheaper than the other commonly used zeolites such as siliceous and aluminous zeolites which were very costlier than the sodium zeolite. Therefore, sodium zeolite can be used as cost effective adsorbent for the removal of phenol from aqueous solution.

The objective of this study was to make use of the sodium zeolite as an adsorbent for adsorption of phenol, to characterize it and to investigate the effects of initial phenol concentration, contact time, pH and adsorbent dosage on the adsorption of phenol on sodium zeolite. Equilibrium isotherm data were fitted to the Langmuir and Freundlich equations.

MATERIALS AND METHODS

Experimental Procedure

A Commercially available sodium zeolite was used for these experiments and all the impurities and unwanted materials were removed as the adsorption. The adsorption of phenol on sodium zeolite was carried out using batch techniques at room temperature.

Effect of pH

The effect of pH on the amount of phenol removal was analyzed over the pH range from 3 to 9. In this study, 100ml of phenol solution of different concentrations (100ppm, 200ppm, 300ppm, 400ppm, 500ppm) was taken in a stopper flask and was agitated with about 2.0g of sodium zeolite using a shaker. Agitation is made for about 2hr at constant oscillation of about 150osc/min. The pH is varied for each and every sample separately. The samples were then filtered using a filter paper and analyzed.

Contact time



About 100ml of the phenol solution having concentration of about 100ppm is taken in a stopper flask. About six samples of same concentration were taken and agitated with about 2.0g of the adsorbent. Agitation was carried out at a constant oscillation of about 150osc/min. The agitation of each sample was carried out for about 10, 20, 30, 40, 50, 60min respectively. Then the sample was filtered and then analyzed. The above experiment is repeated for different concentrations (200, 300, 400, 500ppm)

Effect of adsorbent dosage

The effect of the sodium zeolite on the amount of removal of phenol solution was obtained by contacting 100ml of the phenol solution of initial phenol concentration of 100ppm with different weight of sodium zeolite in a stopper conical flask. Each sample was then agitated for about 40min at a constant oscillation of about 150osc/min. The samples were then filtered and then analyzed. The above experiment is repeated for different phenol concentrations and adsorbent amounts.

Adsorption Equilibrium

The equilibrium studies is carried out by contacting 0.2 to 2g of the adsorbent with 100ml of phenol solution of different initial concentrations (100ppm, 200ppm, 300ppm, 400ppm, 500ppm) in stopper conical flasks. The samples were then shaken at a constant oscillation of 150osc/min for about 40min in a shaker. After the equilibrium is achieved, the concentrations of the samples were then analyzed.

RESULTS AND DISCUSSION

Effect of pH

The initial pH of adsorption medium is one of the most important parameters affecting the adsorption process. The Figure 1 shows the effect of pH on the adsorption of phenol. It was observed that the uptake of the phenol by sodium zeolite was almost constant in the pH range of 3-6. Then the adsorption decreased with the increasing pH and it decreased from pH 7.0 to pH9. So an optimum pH of about 5.0 is taken for all the other experiments.

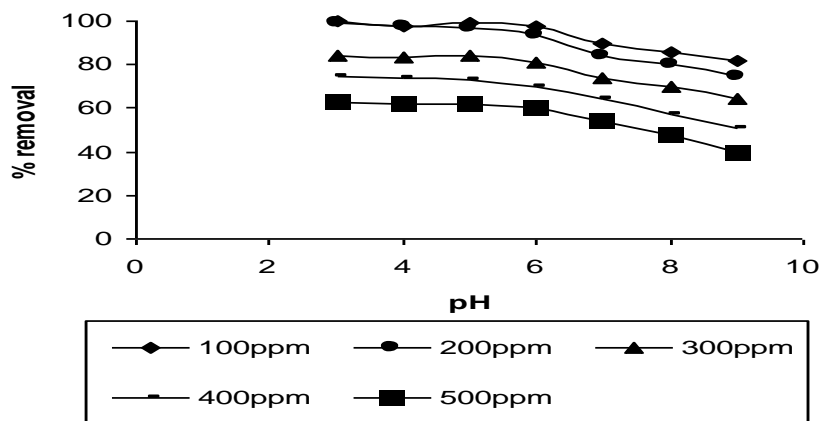


Figure 1 Effect of pH on the adsorption of phenol

Contact time

The result of the contact time experiments is shown in the Figure 2. From the figure, it is clear that the optimum contact time for the experiment to be carried out is about 40 min. above which the equilibrium is achieved and the adsorption remained constant above 40 min.

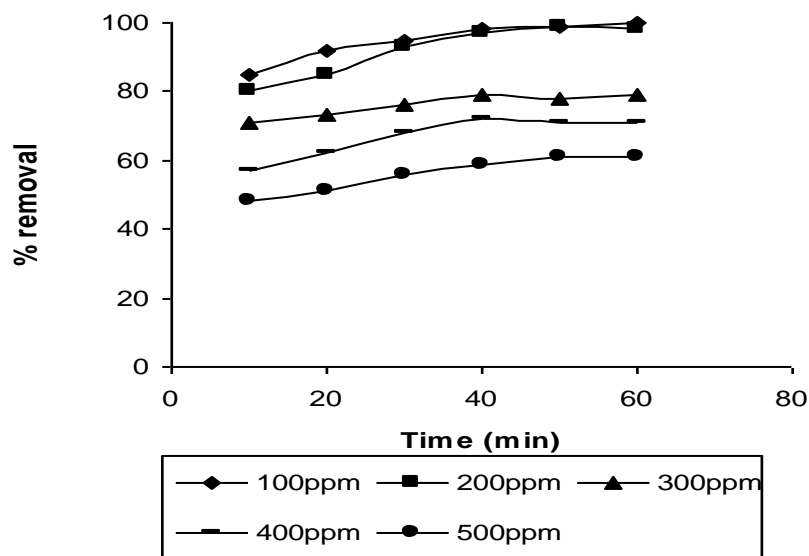


Figure 2 Effect of contact time on the phenol adsorption

Effect of adsorption dosage

To investigate the effect of the mass of the adsorbent on the adsorption of phenol, a series of adsorption experiments was carried out with different adsorbent dosage at an initial

phenol concentration of 100mg/L. The percentage removal of the phenol increased with increasing adsorbent dosage. This can be attributed to increased adsorbent surface area and availability of more adsorption sites resulting from the increase adsorbent dosage as per Table 1.

Table 1 Effect of adsorption dosage

ppm	Adsorbent dosage (g)	% removal
100	0.8	96
	1.0	100
	1.2	100
200	1.8	93
	2.0	100
	2.2	99
300	2.8	96
	3.0	100
	3.2	100
400	3.8	94
	4.0	100
	4.2	99
500	4.8	96
	5.0	100
	5.2	100

Adsorption equilibrium

Equilibrium study on adsorption provides information on the capacity of the adsorbent. The study of adsorption was carried out using isotherm models [9, 12] isotherm models using standard procedures used various authors [18, 2, 15] which provide the basis for the design of adsorption systems. The most widely used isotherm equation for modeling of the adsorption data is the Langmuir equation, which is valid for monolayer sorption onto a surface with a finite number identical site and is given by equation below,

$$Q = \frac{qkC}{1+kC}$$

Where q and k are Langmuir parameters related to maximum adsorption capacity and free energy of adsorption, respectively. C is the equilibrium concentration in the aqueous solution and q is the equilibrium adsorption capacity of adsorbent. The linearized form of Langmuir equation can be written as follows,

$$\frac{C}{Q} = \frac{C}{q} + \frac{1}{qk}$$

The Langmuir constant q and k can be calculated by plotting (C/Q) versus (C) .

The Freundlich model is an empirical equation based on adsorption on heterogeneous surface. It is given as,

$$Q=K(C^{1/n})$$

Where K and n are the Freundlich constants that indicate adsorption capacity and adsorption intensity, respectively.

The linearized form of Freundlich isotherm can be written as,

$$\ln Q = \ln K + (1/n) \ln C$$

The value of K and n can be calculated by plotting $\ln Q$ versus $\ln C$.

Figure 3 and Figure 4 shows the Langmuir and Freundlich curves for phenol adsorption onto sodium zeolite, respectively. It was observed that the equilibrium data were very well represented by the Langmuir isotherm equation when compared to the Freundlich equation. The adsorption data fitted Langmuir and Freundlich equations with correlation coefficients value of 0.9988 & 0.9440 respectively. The best fit of equilibrium data in the Langmuir isotherm expression predicted the monolayer coverage of phenol onto sodium zeolite from above table it was also observed that the maximum adsorption capacity of sodium zeolite was found to be 13.051mg/L.

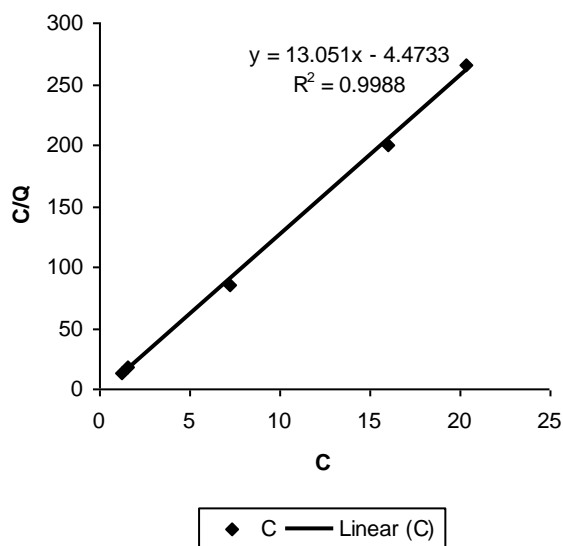


Figure 3: Langmuir isotherm

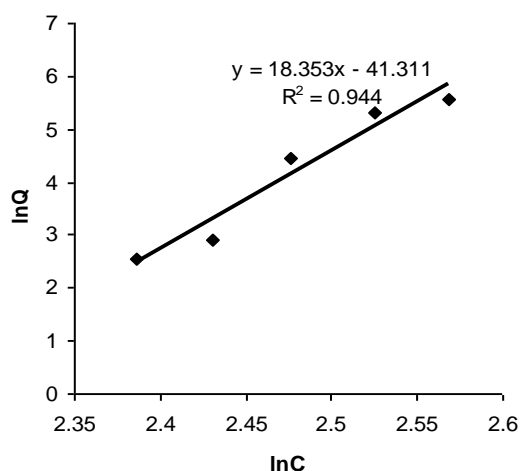


Figure 4: Freundlich isotherm

CONCLUSION

In this study adsorption of phenol from aqueous solution was investigated using sodium zeolite as an adsorbent. The results indicated that adsorption capacity of the adsorbent was considerably affected by initial pH, contact time, and adsorbent dosage. The result also indicated that the uptake of phenol took place at a pH range of 3-6. Then the adsorption of phenol decreased with increasing pH. Equilibrium data fitted well in a Langmuir isotherm equation, confirming the monolayer sorption capacity of phenol onto sodium zeolite with a monolayer sorption capacity of 13.051 mg/L.

REFERENCES

- [1] Agency for Toxic Substances and Disease Registry (ATSDR). Toxicological profile for phenol. Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service. 1998.
- [2] Aksu Z, Yener J. Waste management 2001; 21: 695-702.
- [3] Asheh SA, Banat F, Aitah LA. Sep and Purification Tech 2003; 33:1-10.
- [4] Banat, Al-Bashir FA, Al-Asheh S, Hayajneh O. Environ Pollution 2000; 107: 391-398.
- [5] Bekkouche S, Bouhelassa M, Salah NH, Meghlaoui FZ. Desalination. 2004; 166: 355-362.
- [6] Dursun G, Cicek H, Dursun AY. J Hazardous materials 2005; B125: 175-182.
- [7] Dutta NN, Patil GS, Brothakur S. Sep Sci and Tech 1992; 27: 1435.
- [8] El-Geundi. Adsorp Sci Technol 1997; 15:777-787.
- [9] Freundlich HMF. J Phy Chem 1906; 57A:385-470.
- [10] Kummar S, Upadhyay SN, Upadhyay YD. J Chem Technol Biotechnol 1987; 37:281.
- [11] 11 A, Pilato LA. Springer-Verlag 1985; 104.



- [12] Langmuir. J Am Chem Soc 1916; 38(11):2221-2295.
- [13] Lanouette KH. Chem Eng 1977; 84(22): 99-106.
- [14] Mahajan SP. Pollution Control in Processes Industries - Tata McGraw-Hill, New Delhi 1994.
- [15] Mahvi AH, Maleki A, Eslami A. Am J Appl Sci 2004; 1(4): 321-326.
- [16] Mostafa MR, Sarma SE, Yousef AM. Ind J Chem 1989; 28A: 94-98.
- [17] Phenol. Chemical Week 2002; 164: 31.
- [18] Rengaraj S, Seuny-Hyeon M, Sivabalan R. Waste Management 2002; 22: 543-548.
- [19] Roostaei N, Tezel FH. J Env Management 2004; 70:157-164.
- [20] Singer PC, Chen Yu Yen. Active Carbon {Adsorption of Organics Phase, Vol. 1. (Eds. I.H. Suffet&M.J.McGuire) Ann Arbor Science Publisher Inc., Michigan 1980: 167.
- [21] Spiker JK, Crawford DL, Thiel EJ. Appl Microbiol Biotechnol 1992; 37: 518-523.
- [22] Srihari V, Das Ashutosh. Asian J Microbio Biotech & Envi Sci 2005; 7(3): 469-472.
- [23] Street M, Patrick JW, Camporro-perez MJ. Water Sci Res 1995; 29: 467.