

Research Journal of Pharmaceutical, Biological and Chemical Sciences

Therapeutic Effect of Camel Milk Against Hepatotoxicity Induced by CCl₄ in Rats.

Gabr SA^{1*}, Zahran F², Mohammed Faten F³, Hassanin WF¹, Sharoud MN¹, and Mesalam NM¹.

¹Biological Applications Department, Nuclear Research Centre, Atomic Energy Authority, Egypt.

*Faculty of Education and Science, Taif University, Saudi Arabia Kingdom

²Biochemistry Department, Faculty of Science, Zagazig University, Egypt

³Pathology Department, Faculty of Veterinary Medicine, Cairo University, Egypt

ABSTRACT

This study was aimed to assess the therapeutic potential of camel milk on the carbon tetrachloride (CCl₄)-induced hepatic injury in rats. A total of 24 Rats were randomized and divided into 4 groups (6 rats for each). Group 1: control untreated, group 2 was orally treated with camel milk (5 ml/ rat/day) through gastric intubation with duration of 3 times weekly for 2 weeks and 5 times weekly for another 4 weeks. Group 3: CCl₄ intoxicated rats (intraperitoneally injected with CCl₄(1ml/kg b.w, 3 times weekly for 4 weeks). Group 4 was treated with CCl₄and camel milk with the same dose and treatment protocol of the group 2 &3 .Blood samples were collected at the end of experiment for determination of serum levels of liver enzymes, albumin and total proteins. Determination of inflammatory cytokines (TNF- α and IL-1 β) in liver homogenate was done and detection of the hepatic m-RNA expression of CYP2E1. Other liver specimens were routinely fixed and processed for histopathological evaluation in addition, immunohistochemical evaluation of α - SMA expression in hepatic sections was performed. Results revealed that CCl₄ induced significant (P<0.01) increase in serum hepatic enzymes and significant (P<0.01) reduction of serum total proteins and albumin with elevated levels of TNF- α and IL-1 β compared with control animals. Genetic results showed that the administration of CCl₄ caused a significant down - regulation of the expression of CYP2E1 genes in liver tissue compared to control. On other hand the treatment with camel milk markedly improve serum hepatic functions and inhibit the down regulation of inflammatory cytokines and CYP2E1 genes in liver .Various histopathological alterations were detected in CCl₄- intoxicated group that was markedly ameliorated by camel milk administration. Conclusion: the present study proved the therapeutic potential of camel milk against CCl₄ - induced hepatic damage though improvement of hepatic function, decrease level of inflammatory cytokines and inhibit the down regulation of CYP2E1 genes in liver as well as markedly reduced hepatic histopathological alterations .

Keywords: CCl₄, Hepatotoxicity, Camel milk, Rats.

*Corresponding author

INTRODUCTION

The liver is pivotal organ in body it has several biochemical, metabolic functions, and energy production (Asija et al., 2015). Liver disease is considered as one of most common diseases among humans at different ages and causes great mortalities among public as 20,000 deaths occur every year (Pramod et al., 2012). Viral hepatitis, alcohol liver disease, non-alcoholic fatty liver disease, autoimmune liver disease, metabolic liver disease, drug induced liver injury, gallstones are the most encountered hepatic disorders. Hepatocellular carcinoma is one of the ten most common tumors (Hurkadaleet al., 2012).

Carbon tetra chloride (CCl_4) was known to be hepatotoxin that induced liver damage (Gnanaprakasheta., 2010). Within the body, CCl_4 breaks down to highly toxic trichloromethyl and trichloromethyl peroxy free radicals by cytochrome P450 enzyme (Khan et al., 2009) which cause damage to hepatocytes (Girish et al., 2009). CCl_4 -induced hepatotoxicity via activation of the toxicant by the microsomal cytochrome P450-dependent monooxygenase system to the trichloromethyl radical (CCl_3) and in the presence of oxygen will be converted to a peroxy radical ($\text{CCl}_3\text{-OO}$). These free radicals further induced lipid peroxidation and degenerative cellular changes damage (Boll et al., 2001). These processes are followed by release of inflammatory cytokines and growth factors and depletion of CYP2E1 activity the infiltration of inflammatory cells and release of various cytokines and growth factors (Simeonova et al., 2001, Fahmy et al., 2009, Al-Seeniet al., 2016).

Current available drug treatment is unable to meet the demand clinically due to lack of complete cure, numerous adverse effects, lower safety, etc. Therefore, interest concerning the use of alternative medicines for treatment of hepatic disease has been arisen. Camel milk (CM) is different from other ruminant milk; it has low levels of protein, cholesterol and sugar, but has high levels of vitamins, minerals, and insulin. (Yousef, 2004). It has no allergic features and can be used by lactose-intolerant persons as well (Cardoso et al., 2010). Additionally, CM exhibits a wide range of biological activities; antioxidative, antimicrobial, antihypertensive, antithrombotic, and immuno-modulatory effect (FitzGerald and Meisel, 2000 and Saltanat et al., 2009).

The present study aimed to evaluate the camel milk as a therapeutic agent against CCl_4 -induced hepatic injury.

MATERIALS AND METHODS

Chemicals:

Carbontetrachloride (CCl_4) is of molecular weight 153.84. It was obtained from Loba Chemie, India.

Materials:

Camel milk: Early morning, hand milking CM samples were daily collected from western desert camel farm in sterile screw capped containers and were transported to the laboratory in cool boxes. CM was given to rats in a dose of 5 ml/ rats according to (El Miniawy et al., 2014), once daily, 3 times weekly for 2 weeks and 5 times weekly for 4 weeks more.

Experimental Animals and design

Twenty four adult male albino rats with average body weight 180 gm. Animals were maintained in the animal holding room under controlled environmental conditions (12/12 h light/dark cycle and 24°C) and fed rodent diet (NRC, 1977) and tap water *ad libitum*. They were housed in a well-ventilated vivarium of the animal house of Nuclear Research Centre, Inshas, Egypt. The rats were fed standard diet and water *ad libitum*. The animals were divided into four groups of six animals each and labelled (Groups G1 to G4). Group 1 (G1) served as normal control group. Group 2 (G2) served as camel milk group with oral intubation of rats at a dose of 5 ml / kg once daily, 3 times weekly for 2 weeks and 5 times weekly for 4 weeks more. Group 3 (G3) served as CCl_4 - intoxicated group, intraperitoneally injected (I.P) with CCl_4 (1 ml/kg 3 times weekly for four weeks according to Abdel-Moneim et al. (2015). Group 4 (G4) served as therapeutic camel milk group, the rats were I.P injected

with CCl₄ for 4 weeks, beginning from 3rd week rats were orally treated with CM along with CCl₄ then increasing the times of CM treatment to 5 times weekly for 4 weeks more.

Determination of liver enzymes activities and total bilirubin

Serum samples were used for determination of alanine and aspartate aminotransferase activities (ALT&AST) (Reitman and Frankel, 1957), alkaline phosphatase (ALP) (Belfied and Goldberg, 1971) and total bilirubin (TP) (Walter and Gerade, 1970).

Determination of serum Total protein and albumin.

The level of total proteins and albumin were determined by a colorimetric method as described by Gornalet *al.* (1949) and Doumaset *al.* (1971) respectively using available commercial kits.

Determination of serum globulin (G)

The globulin value was obtained by subtracting the albumin value from the corresponding total proteins value for each sample.

Determination of serum globulin /albumin (A/G) ratio

The A/G ratio was calculated by dividing each sample's albumin value by its corresponding albumin value.

Determination of TNF- α and interleukin -1- beta (ELISA assay):

liver levels of tumour necrosis factor alpha (TNF- α) and interleukin -1- beta (IL-1- β) in rats were measured using enzyme-linked immunosorbent assay (ELISA) kit by the method described by Taylor (2001) and Grassiet *al.* (1991) respectively according to manufacturer's instructions.

Histopathological examination and lesion scoring

Liver specimens were collected from different treated groups and were fixed in 10% buffered formaline. Tissue specimens were routinely processed to obtain paraffin embedded histological section which were routinely stained by H&E stain in addition to special Masson's Trichrome for collagen staining. (Bancroft and Gamble, 2008). Hepatic sections were microscopically examined and lesion scoring was performed as follow:-

Examination of 10 low power fields (4x for fibrosis and 10x for the other hepatic lesions) per liver section and five liver sections per group were examined. Fibrosis was graded according to Ishaket *al.* (1995) into six grades. grade 0 indicates no fibrosis, grade 1 indicates fibrosis of some portal areas with or without short septa expansion into hepatic lobule, grade 2 indicates fibrosis of most portal areas with or without short septa expansion into hepatic lobule, grade 3 indicates fibrosis of most portal areas with occasional portal to portal bridging, grade 4 indicates fibrosis of most portal areas with marked portal to portal bridging and finally grade 5 that indicates fibrosis of most portal areas with marked portal to portal bridging and occasional nodule formation and finally grade six that indicates complete cirrhosis. the confluent necrosis was also graded into six grades according to Ishaket *al.* (1995) as follow :grade 0 for no obvious necrosis, grade 1 indicated focal confluent necrosis, grade 2 indicates Zone 3 necrosis in some areas, grade 3 indicates Zone 3 necrosis in most areas, grade 4 indicates Zone 3 necrosis with occasional bridging portal-central, grade 4 indicates Zone 5 necrosis with multiple bridging portal-central and grade 6 for Panacinar necrosis, while ballooning degeneration, steatosis and hepatocellular vacuolization were graded into four grades according to Shackelford *et al.* 2002). Grade 1 indicated that lesion was present but so minor, grade 2 indicate that lesion was identified but not prominent, grade 3 indicates that lesion was prominent feature and finally grade 4 indicated that lesion was overwhelming feature in the tissue.

Immunohistochemical analysis of α -SMA:

Liver sections were deparaffinization in xylene for 15 minutes, rehydration in graded ethanol, blockage of endogenous peroxidase was done adding few drops of H₂O₂. antigen retrieval was carried out using heat treatment in microwave at 500 W for 10 min by adding 10 mM citrate buffer, pH 6.0 over the slide and put the slides in the microwave. Sections were incubated overnight at 4 °C in a humidified chamber with one of the following primary antibodies: mouse monoclonal antibody to α -SMA diluted 1:100(mouse anti- α -SMA, clone 1A4, DAKO). Anti-mouse IgG in rabbit (cat no. M7023; 1:500; Sigma-Aldrich) was used as the secondary antibody. The sections were washed with PBS. sections were incubated with Streptavidin peroxidase (Thermo Scientific). slides were incubated for 10 min with 3,3'-diaminobenzidine tetrahydrochloride (DAB, Sigma). Finally, the slides were counterstained with haematoxylin then dehydrated and mounted. The cells that were stained brown staining in the cytoplasm/nucleus were considered to be positive.

For immune his to chemical examination, anti- α -SMA (Santa Cruz, CA, USA) was used as primary antibodies to detect its targeted protein using standard immune his to chemical method. Stained tissues were determined using a light microscope (Olympus BX 51, Olympus America, Melville, NY) and photographed with a digital camera (Olympus DP11) connected to the microscope.

Gene expression assay

Semi-quantitative RT-PCR

Isolation of Total RNA and Real-Time PCR (qPCR)

Total RNA was purified from 30 mg of liver tissue using Qiagen tissue extraction kit (Qiagen) according to the manufacturer's protocol. The purity (A260/A280 ratio) and the concentration of RNA were obtained using spectrophotometry (dual wave length Beckman, Spectrophotometer, USA). GAPDH was used as a housekeeping gene for normalizing mRNA levels of the target genes. The mRNA expression levels of CYP2E1 gene was assessed using qPCR standardized by co-amplification with the housekeeping gene GAPDH. Briefly, the total RNA was reverse transcribed into cDNA by reverse transcriptase kit (Fermentas, USA). cDNA was added to a Quantifast SYBR Green qPCR Master Mix (Qiagen) containing 3 μ l of each primer (Table 1).

Table 1: Primers sequences

Gene	Primer Sequence	Reference
CYP2E1	F: 5'- TCCAGGTTTGCACCAGACTCT-3' R: 5'- TCCTCGCTCCTCCTGAGAAG-3'	(Galalet <i>al.</i> , 2014)
GAPDH	F: 5'-ACCACAGTCCATGCCATCAC-3' R: 5'-TCCACCACCCTGTTGCTGTA-3'	(Ogalyet <i>al.</i> , 2015)

The thermal profile included 40 cycles of denaturation at 95 °C for 15 s, annealing at 60 °C for 15 s and extension at 72 °C for 45 s. During the first cycle, the 95 °C step was extended to 1 min. The GAPDH gene was amplified in the same reaction to serve as the reference gene. Gene expression levels were calculated and determined following the method described byLivak and Schmittgen. (2001).

Statistical analysis:

Statistical differences between the means were assessed by analysis of variance (ANOVA) according to Snedecor and Cochran (1982) followed by Duncan's multiple range test(Duncan, 1955)using(SPSS for Windows, version 19). Lesion scoring was done using T- test. Values of P<0.05 were considered statistically significant.

**RESULTS**

Table (2) showed insignificant difference in the mean values of serum total protein, albumin, AST, ALT, ALP, total bilirubin between normal control group (G 1) and CM- treated group (G3).

Table (2) Effect of camel milk treatment the activities of alanine transaminase (ALT), aspartate transaminase (AST), alkaline phosphatase (ALP) enzymes, total bilirubin and serum protein fractions of normal rats and ccl4 - intoxicated ones.

Parameters Group	T. Protein (g/dl)	Albumin (g/dl)	Globulin	A/G ratio	T. Bilirubin (mg/dl)	AST (U/ml)	ALT (U/ml)	ALP (U/ml)
Normal control	8.38 ^a ±0.15	4.46 ^a ±0.04	5.27 ^a ±0.19	0.85 ^b ±0.04	0.89 ^c ±0.02	164.17 ^b ± 0.91	63.33 ^b ±0.84	193.21 ^b ±37.07
CCl ₄	5.42 ^b ±0.26	3.17 ^b ±0.21	2.25 ^c ±0.08	1.41 ^a ±0.0.08	3.30 ^a ±0.11	203.43 ^a ±4.96	173.71 ^a ±13.89	464.42 ^a ±9.03
CM	8.56 ^a ±0.17	4.39 ^a ±0.18	4.17 ^b ±0.08	1.06 ^b ±0.06	0.79 ^b ±0.02	172.33 ^b ±7.40	70.17 ^b ±2.01	173.57 ^b ±2.54
CCl ₄ +CM	9.04 ^a ±0.42	4.23 ^a ±0.11	4.81 ^{ab} ±0.45	0.92 ^b ±0.09	1.26 ^c ±0.0	168.00 ^b ±1.79	75.83 ^b ±2.39	180.65 ^b ±0.74

Data are expressed as Mean ± S.E

^{a,b,c} Mean values with different letters in the same column are significantly different at (P< 0.01)

CCl₄ treatment of the rats resulted in a significant elevation (P < 0.01) in the mean values of liver functions (AST, ALT and ALP), A/G ratio and total bilirubin. Meanwhile, the mean values of total protein, albumin and globulin were significantly decreased (P < 0.01) compared to control rats. Concomitant oral administration of camel milk with CCl₄ showed significant decrease (P < 0.01) in the mean values of liver enzymes and total bilirubin and the mean values of total protein, albumin and globulin were significantly increased as compared with CCl₄-treated rats alone (G2).

Effects of camel milk on the levels of proinflammatory mediators (hepatic IL-1β and TNF-α Levels).

It was evident from table (3) that CCl₄ intoxicated rats showed elevated levels of proinflammatory mediators, including TNF-α and IL-1β in the liver tissue that were significantly increased when compared with the control group (p<0.01), suggesting induction of a severe inflammatory response. Nevertheless, camel milk (G4) markedly inhibited the levels of these proinflammatory mediators compared to CCl₄- intoxicated group (G2).

Table 3: Effect of camel milk treatment on the values of hepatic IL-1β and TNF-α of normal rats and CCl₄ – intoxicated ones.

Parameters Group	IL-1β	TNF-α
Normal control	32.55 ^c ±1.15	31.43 ^c ±1.68
CCl ₄	127.41 ^a ±4.53	164.64 ^a ±9.42
CM	27.58 ^c ±0.42	30.57 ^c ±0.87
CCl ₄ +CM	56.57 ^b ±3.85	58.38 ^b ±1.09

Data are expressed as Mean ± S.E

^{a,b,c}Mean values with different letters in the same column are significantly different.

Gene Expression Analysis:

Real-time quantitative PCR analysis showed a significant decrease in CYP2E1 mRNA level in CCl₄-intoxicated group compared to that of control rats. However, CM treatment to CCl₄-intoxicated rats (G4) induced a markedly attenuated CYP2E1 down-regulation by increasing its mRNA levels significantly compared to the CCl₄ group (G2). No significant difference between CCl₄ group (G2) and normal control ones (Table 4).

Table 4: Effect of camel milk treatment on the values of CYP 2E1 of normal rats and CCl₄ – intoxicated ones.

Group parameter	Normal control	CCl ₄	CM	CCl ₄ +CM
CYP 2 E1	1.09 ^a ±0.03	0.17 ^b ±0.05	1.01 ^a ±0.02	1.04 ^a ±0.01

Data are expressed as Mean ± S.E

^{a,b}Mean values with different letters in the same column are significantly different.

Effect of camel milk on CCl₄-induced liver histopathological alternations and lesion scoring:

The results of hepatic histopathological lesion scoring of different organs were shown in Table (5) when compared with the normal liver tissues of controls, liver tissue in the rats treated with CCl₄ (G2) revealed extensive liver injuries, characterized by cholangiofibrosis represented by portal fibrosis, oval cell proliferation with formation of bile ductules (fig 1a) and macro and micro vesicular steatosis of hepatocytes associated with hepatocellular necrosis of periportal hepatocytes (Fig 1b). However, the histopathological hepatic lesions induced by I.P injection of CCl₄ were remarkably ameliorated by treatment camel milk. This finding was consistent with the levels of the enzymes markers. These effects were markedly rehabilitated by CM. The liver

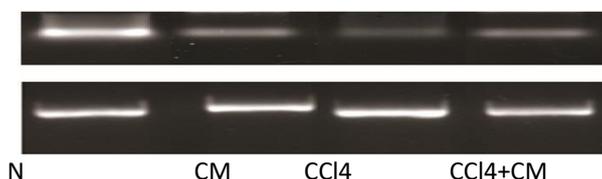
specimens obtained from the rats intoxicated with CCl₄ and post- treated with CM resulted in reduction in the severity of hepatocellular lesions induced by CCl₄, lesions were restricted to c) minimal portal fibrosis with maintaining of hepatic lobular structure and vacuolization of hepatocellular cytoplasm.(fig 1c) and Minimal portal mononuclear cell infiltration and vacuolization of periportal hepatocytes (fig 1d).Concerning hepatic lesion in camel milk – treated rats was restricted to showing diffuse vacuolization of hepatocytes (fig 1e).Rat of untreated control group showing normal histological hepatic structure (fig 1f).

Table 5: Histological injury score of liver intoxicated with CCl₄ and treated with camel milk.

Score injury group	Hepatic fibrosis	Confluent necrosis	Ballooning degeneration	Macro to microvesicula rsteatosis
CCl ₄	2.82 ^a ±0.21	3.30 ^a ±0.81	1.65 ^a ±0.14	2.13 ^a ±0.32
CCl ₄ +CM	1.42 ^b ±0.10	1.92 ^b ±0.16	0.00 ^b ±0.00	0.00 ^b ±0.00

Data are expressed as Mean ± S.E.

^{a,b}Mean values with different letters in the same column are significantly different.



His to chemical and immune his to chemical findings

Masson’s trichrome staining of the liver was performed to assess collagen fiber distribution.The histochemical staining of hepatic section by Masson’s Trichrome revealed marked variation in fibrosis among CCl₄ treated groups. In CCl₄ group there was bluish stained collagenous tissue disrupting the hepatic parenchyma note the massive bridging fibrosis with pseudolobules formation (Masson’s Trichrome,X100) (fig 2a). Liver of rat from CCl₄+CM treated group showing bridiging fibrosis grade 4 note the collagenous tissue proliferation extending from portal to portal and portal to central (Masson’s Trichrome,X100). (fig 2b). On other hand Liver of CCl₄ treated rat showing positive stained cells are spindle to stellate with large amount of cytoplasm and long extending cytoplasmic processes (Immunohistochemistry for alpha-SMA, X400). (fig 2c). Liver of rat from CCl₄+CM treated group showing many brownish staining immune positive cells in the area of portal fibrosis (Immunohistochemistry for alpha-SMA,X400).(fig 2d).

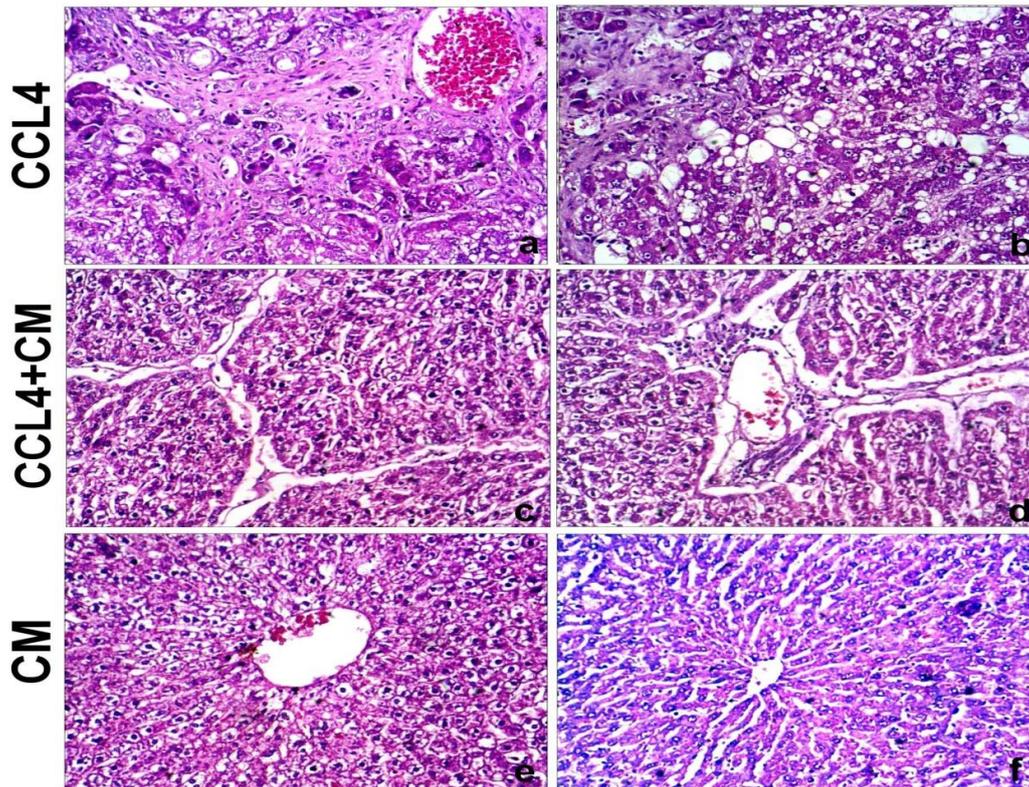


Fig 1: liver of rat from: CCl₄- intoxicated group showing: a) cholangiofibrosis represented by portal fibrosis, oval cell proliferation with formation of bile ductules and b) macro and microvesicular steatosis of hepatocytes associated with hepatocellular necrosis of periportal hepatocytes. Liver of rat from CCl₄ - intoxicated group plus CM showing: c) minimal portal fibrosis with maintaining of hepatic lobular structure and vacuolization of hepatocellular cytoplasm. d) Minimal portal mononuclear cell infiltration and vacuolization of periportal hepatocytes. e) rat from CM treated group showing diffuse vacuolization of hepatocytes. f) rat of untreated control group showing normal histological hepatic structure (H&E, X200).

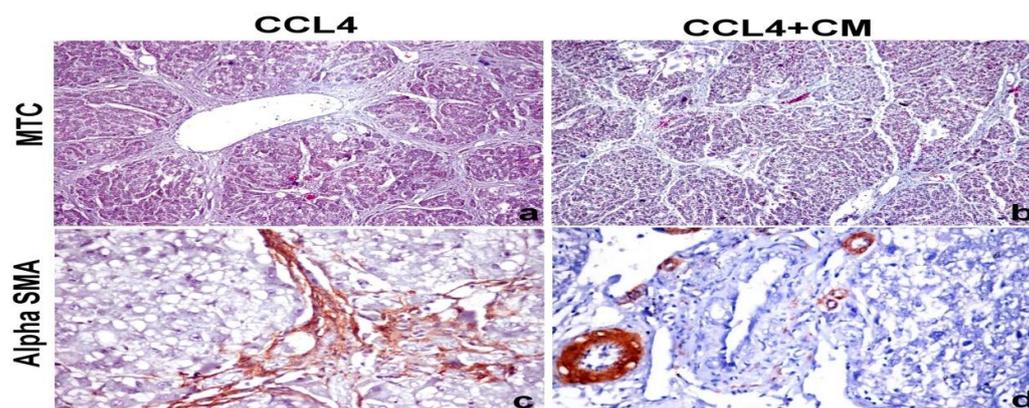


Fig 2: a) Liver of CCl₄ treated rat showing bluish stained collagenous tissue disrupting the hepatic parenchyma note the massive bridging fibrosis with pseudolobules formation (Masson's Trichrome, X100). b) liver of rat from CCl₄+CM treated group showing bridging fibrosis grade 4 note the collagenous tissue proliferation extending from portal to portal and portal to central (Masson's Trichrome, X100). c) Liver of CCl₄ treated rat showing positive stained cells are spindle to stellate with large amount of cytoplasm and long extending cytoplasmic processes (Immunohistochemistry for alpha-SMA, X400). d) liver of rat from CCl₄+CM treated group showing many brownish staining immune positive cells in the area of portal fibrosis (Immunohistochemistry for alpha-SMA, X400).

DISCUSSION

The present study was aimed to assess the therapeutic effect of camel milk carbon tetrachloride treated rats. It is bio transformed by the cytochrome P 450 system. By this process, it produces the trichloromethyl free radical, which in turn covalently binds to cell membranes and organelles and elicits lipid peroxidation. It also disturbs Ca^{2+} homeostasis and finally results in cell death (Recknaget *et al.*, 1989). Increased levels of ALT, AST, ALP, TP, Alb and globulin are conventional indicators of liver injury (Achiliyaet *et al.*, 2004). The activities of serum marker enzymes, like ALT, AST and ALP when estimated can make the assessment of liver function when liver cell plasma membrane is damaged, a variety of enzyme which are normally located in the cytosol are released into the blood stream. Due to the structural damage of liver, the enzyme levels are increased in serum because of their location in cell cytoplasm. After damaging or injury they are released into blood circulation and raises the level of enzymes in serum (Sultana and Nazam, 2012). During hepatic damage, cellular enzymes like AST, ALT and ALP present in liver cells leak into the serum, resulting in increased concentrations (Patel and Jawaid, 2014).

The estimation of serum marker enzymes like ALT, AST and ALP in the serum is a useful quantitative marker of the extent and type of hepatocellular damage (Mitra *et al.*, 1998). In CM injected group, the tendency of these enzymes to return to near normal is a clear manifestation of antihepatotoxic effects of the extract. The reduction in the levels of enzymes like ALT and AST towards the normal value is an indication of regeneration process. Bilirubin, the endogenous product derived from the degradation of haemoglobin, builds up in the blood and extracellular fluid as its excretion is impaired. Usually, ALT and AST or in combination with total bilirubin are estimated for assessment of hepatocellular injury in rodents and non-rodents (Singh *et al.*, 2011). The reduction in ALP levels with concurrent depletion of raised bilirubin levels is suggestive of the stability of the biliary function during injury with carbon tetrachloride. Histopathological liver sections also revealed that the normal liver architecture was disturbed by hepatotoxin, CCl_4 whereas, in the liver sections of the rat treated with the CM showed the normal cellular architecture was retained. Several studies revealed that CCl_4 treatment increased levels of AST, ALT, ALP and bilirubin. (Ahsan *et al.*, 2009, Soma *et al.*, 2014 and Yengkhomet *et al.*, 2017) and Similarly, in present study there was significant rise of ALT, AST, ALP and bilirubin levels in the CCl_4 treated groups signifying the induced liver injury.

The levels of albumin and total proteins were reduced due to the CCl_4 - induced hepatotoxicity. The reduction is attributed to the initial damage produced and localized in the ER (endoplasmic reticulum) which results in the loss of P450 leading to its functional failure with a reduction in protein synthesis. The albumin and protein levels were also raised suggesting the stabilization of ER (endoplasmic reticulum) leading to synthesis of protein. While, the biological value of bilirubin has been employed to assess the excretory role of the liver (Tietz, 1995), the metabolic alterations in the serum concentrations of albumin and total protein are used to monitor its secretory capability (Oloyede and Sunmonu, 2009). In the present study, the significantly increased serum level of bilirubin in the untreated hepatotoxic rats could be associated with CCl_4 -mediated defect in the carrier-mediated saturable system at the sinusoidal surface of the hepatocytes that consequently obstruct bilirubin uptake and secretion into bile (Sabiet *et al.*, 2014). Similarly, the CCl_4 -mediated significant reduction in the levels of albumin and total protein may be suggestive of diminished synthetic function of the liver (Sabiet *et al.*, 2015). CCl_4 -intoxication leads to hypomethylation of cellular components in the case of RNA the outcome is thought to be inhibition of protein synthesis. Hypoproteinemia and hypoalbuminemia in rats intoxicated with CCl_4 for 6 weeks have been reported by Al-Yahya *et al.* (2013). The amelioration of rising serum enzymes in CCl_4 toxicity by CM may be due to the prevention of the leakage of intracellular enzymes by its membrane stabilizing activity. This is in agreement with the commonly accepted view that serum transaminases levels return to normal with the healing of hepatic parenchyma and the regeneration of hepatocytes (Bessem and Vermeulen, 2001 and Darwish *et al.*, 2012). Several studies (Al-Fartosi *et al.*, 2011, Dallak, 2009 and Al-Hashem, 2009) have provided an abundant support for evidencing the protective effects of camel milk on liver damage. The mechanism by which CM lowered liver enzymes may be referred to their ability to maintain liver cell integrity (Ibrahim *et al.*, 2017).

The administration of CM resulted in an increase in total protein and stimulation of protein synthesis has been advanced as a contributory hepatoprotective mechanism which accelerates the regeneration process and the production of liver cells (Rip *et al.*, 1985; Tadeusz *et al.*, 2001).

Furthermore, the histopathological findings of liver samples are in agreement with the results of biochemical studies. CCl₄ caused damage to the hepatic architecture and produced histological changes such as inflammatory cell infiltration, necrosis of hepatocytes and sinusoidal dilatation. These results are in accordance with those obtained by Hsouna *ET AL.*, (2011) and Kale *ET AL.*, (2012) and Laouar *et al.*, (2017) which indicate that CCl₄ cause histopathological liver changes in rats. Liversections of CCl₄treated rats were characterized by significant intracellular lipidaccumulation, ballooning of hepatocytes, infiltration with inflammatory cells and hepatocyte necrosis.(Desai *et al.*, 2012).Moreover, histopathological evaluation of livers revealed that camel milk reduced inflammation, necrosis and the number of liver lesions induced by CCl₄. The above results suggest that Camel milk inhibits CCl₄-induced oxidative hepatic damage by protecting cells from the effects of CCl₄ and by reducing insidious progressive inflammation-induced liver injury.

Since CYP2E1 is the major isozyme involved in the metabolism of CCl₄, the expression of CYP2E1 was investigated. ROS formed during the biotransformation process of CCl₄ are more reactive and toxic than the parental compound. Biotransformation of CCl₄ occurs in the endoplasmicreticulum and the isoenzyme implicated in this process is CYP2E1.As expected, CCl₄ induced a significant reduction of CYP2E1mRNA level (Figure). This specific CYP2E1 dysregulation in CCl₄ hepaticinjury was previously reported by Sakret *al.*(2011) and may result from a direct attack of reactive CCl₄metabolites leading to CYP2E1 transcript degradation. Moreover, other mechanisms such as an inhibitionof CYP2E1 transcription subsequent to inflammatory responses could also have a role in CYP2E1 mRNA decrease(Riddick *et al.*, 2004). An interesting observation in our study was the total blockade of CCl₄-inducedCYP2E1 downregulation by camel milk treatment . These data suggest that the hepatoprotective effect of camel milk possibly due to prevention of CCl₄-induced CYP2E1 downregulation.Chen *et al.*, (2017) found that expression of CYP2E1 protein was decreased after CCl₄ treatment. Also, Mohamed, (2017) showed that cytochrome 2E1 was down regulated when 1ml /kg ccl4 30% in olive oil were given every 72h for 10 days.

HSCs with α -smooth muscle actin (α -SMA) expression play a key role in pathophysiological mechanism of hepatic fibrosis. Thus, α -SMA is a reliable unique marker of fibrosis and could be useful in monitoring the efficacy of the antifibrotic therapy(Carpinoet *al.*, 2005, Domitrovićet *al.*, 2009 and Parikh *et al.*, 2014). Hepatic stellate cells are regarded as the most relevant cell for the development of liver fibrosis, and their activation is the key step in the process of liver fibrogenesis(Tsukamoto, 2005). α -SMA, the marker of activated HSCs following liver injury, were used to evaluate the degree of HSC activation by immune his to chemical staining.CCl₄ treatment significantly increased the accumulation of α -SMA in this study confirming that CCl₄ stimulated the activation of HSCs in the rat model and agreed with(Rockeyet *al.*, 2013). Camel milk significantly improved the liver histology and resolved the fibrotic changes induced by CCl₄ and decreased its progression with a marked reduction of in α -SMA immune reactivity in hepatocytes.

Shim *et al.* (2010) found that inflammatoryytokines, such as TNF- α and IL-1 β were markedly induced in CCl₄-treated mice. TNF- α is a pleiotropic proinflammatory cytokine mainly produced by activated macrophages and monocytes and is involved in many different biological and pathologic processes including inflammation, autoimmune diseases and cancer. In the current study, we found that the levels of proinflammatory mediators, including TNF- β and IL-1 β evidently increased in the liver tissue of rats treated with CCl₄, 3times weekly for 4 weeks, but administration of camel's milk markedly decreased their level. Tan *et al.*(2016)recorded significant elevation ofpro-inflammatory cytokines IL-6, TNF- α , and IL-1 β with hepatic fibrosis in mice treated with CCl₄. In the same line, Ahnet *al.*(2016) and El-Boshyet *al.*(2017)observed elevated pro-inflammatory cytokines, including TNF- α and IL-1 β ,mRNA expression with hepatic damage in rats treated with CCl₄.These results suggest that CM inhibit the hepatic local inflammatory response due to the fact that lactoferrin, an anti-inflammatory protein component of CM, has been reported to inhibit the production of pro inflammatory cytokines e.g., TNF-a in mononuclear cells in vitro and in vivo, in response to lipopolysaccharide activation. Mechanistically, the inhibition of the sepro-inflammatory cytokines production could result from inhibition of NF- κ B activation following internalization of Lactoferrin into monocytes (Haversenet *al.*, 2002).Arabetal. (2014) found that Feeding with CM decreased the levels of TNF-a along with IL-10 in the colons of animals with TNBS colitis. These results imply that CM partly exerted its beneficial effects on colon inflammation by lowering the colonic content of proinflammatory cytokines such as TNF-a. In fact, CM has been reported to suppress inflammation and elevated levels of TNF-a in ethanol-induced hepatic injury (Darwish *et al.*, 2012).Camel milk treatment caused significant decrease in IL-1b, Besides, camel milk treatment caused significant decrease in the TNF-a level (Alhaider *et al.*, 2013).

CM prevents oxidative injury and cell damage by several mechanisms, including scavenging free radicals and inhibiting lipid peroxidation (Althnaian, 2012). The protective effect of camel milk against APAP-induced toxicity, oxidative stress, and tissue damage in this study could be referred to its composition of high levels of vitamins C, A, B2 and E and very rich in magnesium and other trace elements (Barbagallo et al., 1999). These vitamins are antioxidants that are found to be beneficial in preventing the tissues injury caused by toxic agent. Magnesium protects the cell against oxy-radical damage and assists in the absorption and metabolism of vitamins B, C and E (Majerus et al., 1971) which are antioxidants important in cell protection.

Also, CM is rich in zinc (Zn) (Althnaian, 2012). Zinc is a trace element fundamental for living organisms. Many enzymes require Zn for their activity. It also plays a vital role in the DNA replication, transcription, and protein synthesis, affecting cell division and differentiation. (Frederickso, 1989). It has been documented that Zn has a link with many of body enzymes and can prevent cell injury through activation of the antioxidant system (Ozturk et al., 2003 and Ozdemir and Inanc, 2005). Based on the present results, the ability of camel milk to reverse of the severe alterations in the liver injury markers caused by CCl₄ is a clear indication of the improvement of the functional status of hepatocytes with preservation of cellular architecture.

CONCLUSIONS

In light of all findings, the present study suggest that CM can evidently served as anti hepatotoxic agent, which might be related with ameliorating liver functions, improving histopathological alternations, reducing levels of proinflammatory mediators, inhibiting α -SMA production and collagen production and up-regulating CYP2E1 expression. CM might act as a promising complementary treatment to combat hepatotoxicity.

REFERENCES

- [1] Arab, H.H.; Salama S.A.; Eid, A.H.; Hany A.; Omar, H.A.; . Arafa, E.A.; Maghrabi, I.A. (2014): Camel's milk ameliorates TNBS-induced colitis in rats via downregulation of inflammatory cytokines and oxidative stress. *Food and Chemical Toxicology*, 69: 294–302.
- [2] Achiliya, G.S.; Wadodkar, S.O.; Dorle, A.K. (2004): Evaluation of hepatoprotective effect of Amakadi Ghrita against carbon tetrachloride induced hepatic damage in rats. *J. Ethnopharmacol.* 90, 229-232.
- [3] Ahsan, R.; Islam, K.M.; Musaddik, A.; Haque, E. (2009): Hepatoprotective activity of methanol extract of some medicinal plants against carbon tetrachloride induced hepatotoxicity in albino rats. *Global J Pharmacol.* 3(3):116-22.
- [4] Al-Fartosi, K.G.; Khuon, O.S.; Al-Tae, H.I. (2011): Protective role of camel's milk against paracetamol induced hepatotoxicity in male rats. *Int J Res Pharmaceut Biomed Sci.* 2:1795-9.
- [5] Al-Hashem, F. (2009): Camel's milk protects against aluminum chloride-induced toxicity in the liver and kidney of white albino rats. *Am J Biochem Biotechnol.* 5:98-109.
- [6] Althnaian, T. (2012): Protective Effect of Camel Milk Against Carbon Tetrachloride Hepatotoxicity in Rats. *Global Veterinaria* 9 (5): 564-570.
- [7] Asija, R.; Kumar, V.; Sharma, A.K.; (2015): Hepatoprotective Activity of *Lantana Camera* against Carbon tetra Chloride Induced Hepatotoxicity in Wister Rat. *International Journal of Pharmaceutical Erudition*, 4(4): 1-7.
- [8] Bancroft, J. D.; and Gamble, M.; (2008): *Theory and Practice of Histological Techniques*. 6th Ed., Churchill Livingstone, Elsevier, China.
- [9] Barbagallo, M.; Dominguez, L.J.; Tagliamonte, M.R.; Resnick, L. M.; Paolisso, G. (1999): Effects of Vitamin E and Glutathione on Glucose Metabolism Role of Magnesium. *Hypertension*. 34:1002-6.
- [10] Belfield, A.; Goldberg, D. (1971). Colorimetric determination of alkaline phosphatase activity. *Enzyme*, 12: 561-566.
- [11] Bessems, J.G.; Vermeulen, N.P. (2001): Paracetamol (acetaminophen)-induced toxicity: molecular and biochemical mechanisms, analogues and protective approaches. *Crit Rev Toxicol.* 31:55-138.
- [12] Abdel-Moneim, A.M.; Al-Kahtani, M.A.; El-Kersh, M.A.; Al-Omar, M.A. (2015): Free Radical-Scavenging, Anti-Inflammatory/Anti-Fibrotic and Hepatoprotective Actions of Taurine and Silymarin against CCl₄ Induced Rat Liver Damage. *PLoS ONE* 10(12): e0144509. <https://doi.org/10.1371/journal.pone.0144509>.

- [13] Cardoso, R.; Santos, R.; Cardoso, C.; Carvalho, M.(2010)): Consumption of camel's milk by patients intolerant to lactose. A preliminary study. *Review Allergy Mexican*. 57:26-32.
- [14] Carpino, G.; Morini, S.; GinanniCorradini, S.; Franchitto, A.; Merli, M.; Siciliano, M.; Gentili, F.; OnettiMuda, A.; Berloco, P.; Rossi, M.(2005): Alpha-SMA expression in hepatic stellate cells and quantitative analysis of hepatic fibrosis in cirrhosis and in recurrent chronic hepatitis after liver transplantation. *Dig. Liver Dis*. 37, 349–356.
- [15] Chen,Q.; Zhan, Q.; Li, Y.; Sun,S.; Zhao,L.; Zhang, H.; and Zhang, G.(2017): *Schisandra*Lignan Extract Protects against Carbon Tetrachloride-Induced Liver Injury in Mice by Inhibiting Oxidative Stress and Regulating theNF- κ B and JNK Signaling Pathways. *Evidence-Based Complementary and Alternative Medicine*. .1-11.
- [16] Dallak, M.(2009): Camel's milk protects against cadmium chloride-induced hypochromic microcytic anemia and oxidative stress in red blood cells of white albino rats. *Am J PharmacolToxicol*. 2009;4:136-43.
- [17] Darwish, H.A.; AbdRaboh, N.R.; Mahdy, A. (2012). Camel's milk alleviates alcohol-induced liver injury in rats. *Food ChemToxicol*. 50:1377–1383.
- [18] Domitrović, R.; Jakovac, H.; Tomac, J.; Šain, I.(2009): Liver fibrosis in mice induced by carbon tetrachloride and its reversion by luteolin. *Toxicol. Appl. Pharmacol*. 241, 311–321.
- [19] Dumas, B. T.; Watson ,W.A.; and Biggs, H.G. (1971). Albumin standards and the measurement of serum albumin with bromcresol green. *ClinicaChimicaActa* , 31(1):87-96.
- [20] Fahmy SR, Hamdi SAH, Abdel-Salam HA. (2009): Curative effect of dietary freshwater and marine crustacean extracts on carbon tetrachloride-induced nephrotoxicity.*Australian Journal of Basic and Applied Sciences*, 3(3), 2118-2129.
- [21] FITZGERALD RJ, MEISEL H. (2000): MILK PROTEIN-DERIVED PEPTIDE INHIBITORS OF ANGIOTENSIN-I-CONVERTING ENZYME.BR J NUTR. 2000 NOV;84 SUPPL 1:S33-7.
- [22] Frederickson, C.J.(1989): Neurobiology of zinc and zinc-containing neurons. *Int Rev Neurobiol*. 31:145-238.
- [23] Galal, M.K.; Khalaf, A.A.; Ogaly, H.A. and Ibrahim, M.A.(2014): Vitamin E attenuates neurotoxicity induced by deltamethrin in rats. *BMC Complement. Altern. Med*. 14, 458–464.
- [24] Girish, C.; Koner, B.C.; Jayanthi, S.; Rao, K.R.; Rajesh, B. and Pradhan, S.C.(2009): Hepatoprotective activity of six polyherbal formulation in CCl₄ induced liver toxicity in mice. *Indian J ExpBiol*47: 257263.
- [25] Gnanaprakash, K.; Madhusudhana, C.C.; Ramkanth, S.; Alagusundaram, M.; Tiruvengadarajan, V.S. and Angala, P. S.(2010):. Aqueous extract of *Flacourtiaindicap*prevents carbon tetrachloride induced hepatotoxicity in rat. *Int J BiollifeSci*6: 51-55.
- [26] Gornall, A. G.; Bardawill, C. J. and David, M. M. (1949), Determination of serum proteins by means of the biureto reaction. *J. Biol. Chem.*, 177, 751-766.
- [27] Hsouna,A.,B.;Saoudi,M.,Trigui,M.,Jamoussi,K.,Boudawara,T.,Jaoua,S.(2011):Characterization of bioactive compounds and ameliorative effects of CERATONIASILILQUA leaf extract against CCl₄ induced hepatic oxidative damage and renal failure in rats.*FoodChemToxicol*, 49 (12).3183–3191.
- [28] Kale, I.; Khan, M.A.; Irfan,Y. and Goud. V.A. (2012): Hepatoprotective potential of ethanolic and aqueous extract of flowers of *Sesbaniagrandiflora*(Linn) induced by CCl₄, *Asian Pac J Trop Biomed*, 2 (2): S670–S679.
- [29] Khan, M.R.; Rizvi, W.; Khan, G.N.; Khan, R. A andShaheen, S.(2009): Carbon tetrachloride induced nephrotoxicity in rats: protective role of *Digeramuncata*. *J Ethnopharmacol*,; 122: 91-99.
- [30] Laouar, A.; Klibet,F.;Bourogaa, E.;Benamara, A.; Boumendjel, A.;Chefrou, A., Messarah, M. (2017): Potential antioxidant properties and hepatoprotective effects of Juniperusphoenicea berries against CCl₄ induced hepatic damage in rats. *Asian Pacific Journal of Tropical Medicine*, 10(3):263-269.
- [31] Livak, K.J.; Schmittgen, T.D. (2001):Analysis of relative gene expression data using real-time quantitativePCR and the 2⁻ $\Delta\Delta$ CT method. *Methods* 2001, 25, 402–408.
- [32] Majerus, P.W.; Brauner, M.; Smith, M.; Minnich, V. (1971):Glutathione synthesis in human erythrocytes: II. Purification and properties of the enzymes of glutathione biosynthesis. *J Clin Invest*. 1971;50:1637.
- [33] Mohamed, M.A. (2017): Pomegranate ameliorates the inflammatory status and oxidative stress in carbon tetrachloride-induced hepatotoxicity in rats. *RJPBCS* 8(1) 49-56.
- [34] Ogaly, H.A.; Khalaf, A.A.; Ibrahim, M.A.; Galal, M.K. and Abd-Elsalam, R.M.(2015): Influence of green tea extract on oxidative damage and apoptosis induced by deltamethrin in rat brain. *Neurotoxicol. Teratol*.50, 23–31.

- [35] Ozdemir, G. and Inanc F.(2005): Zinc may protect remote ocular injury caused by intestinal ischemia reperfusion in rats. *The Tohoku journal of experimental medicine*. 206:247-51.
- [36] Ozturk, A.; Baltaci, A. K.; Mogulkoc, R.; Oztekin, E.; Sivrikaya, A. and Kurtoglu, E. (2003): Effects of zinc deficiency and supplementation on malondialdehyde and glutathione levels in blood and tissues of rats performing swimming exercise. *Biol Trace Elem Res*. 94:157-66.
- [37] Parikh, J.G.; Kulkarni, A. and Johns, C. (2014): α -Smooth muscle actin-positive fibroblasts correlate with poor survival in hepatocellular carcinoma. *Oncol. Lett.* 7, 573–575.
- [38] Patel, R. and Jawaid, T.(2014): Hepatoprotective activity of aerial parts of plant extract of *Callicarpamacrophyllain* rats. *Pharmacy and Pharmacology Research*,;2(1): 1-8.
- [39] Hurkadale,P.J.; Pournima A Shelar, Siddhalinges G Palled, Yuvaraj D Mandavkar, Ajay S Khedkar.(2012): Hepatoprotective activity of Amorphophalluspaeoniifoliustubersagainst paracetamol-induced liver damage in rats. *Asian Pacific Journal of Tropical Biomedicine*, S238-S242.
- [40] Recknagel, R.O.; Glender, E.A. and Walter, R.L. (1989): Mechanism of Carbon tetrachloride toxicity. *Pharmacology and therapeutics*, 43: 139-54.
- [41] Reitman, S. and Frankel, S. (1957): A colorimetric method for the determination of serum glutamic oxalacetic and glutamic pyruvic transaminases. *Amer. J. Clin. Pathol.*, 28: 56–63.
- [42] Riddick, D.S.; Lee, C.; Bhatena, A.; Timsit, Y.E.; Cheng, P.Y.; Morgan, E.T.; Prough, R.A.; Ripp, S.L.; Miller, K.K. and Jahan, A.(2004):. Transcriptional suppression of cytochrome P450 genes by endogenous and exogenous chemicals. *Drug Metab. Dispos.* 32, 367–375.
- [43] Rip, J.W.; Rugar, C.A.; Ravi, K. and Carroll, K. K. (1985): Distribution, metabolism and function of dolichol and polyprenols. *Prog Lipid Res*, 24:269-309.
- [44] Rockey, D.C.; Weymouth, N. and Shi, Z.(2013): Smooth Muscle α Actin (Acta2) and Myofibroblast Function during Hepatic Wound Healing. *PLoS ONE*, 8, e77166.
- [45] Sakr, S.A.; El-Abd, S.F.; Osman, M.; Kandil, A.M. and Helmy, M.S.(2011): Ameliorative Effect of Aqueous Leave Extract of *Ocimumbasilicum* on CCl4-Induced Hepatotoxicity and Apoptosis in Albino Rats. *J. Am. Sci.* 7, 116–127.
- [46] Saltanat, H.; Li, H.; Xu, Y.; Wang, J.; Liu, F. and Geng, X.(2009): [The influences of camel milk on the immune response of chronic hepatitis B patients]. *Xi baoyu fen zimianyixuezhazhi= Chinese journal of cellular and molecular immunology*. 25:431-3.
- [47] Simeonova, P. P.; Gallucci, R. M.; Hulderman, T.; Wilson, R.; Kommineni, C. Rao, and M. and Luster, M. I. (2001): The role of tumor necrosis factor- α in liver toxicity, inflammation, and liver fibrosis induced by carbon tetrachloride. *Toxicol. Appl. Pharmacol.* 177:112–120.
- [48] Singh, A.; Bhat, T.K. and Sharma, O.P.(2011): Clinical biochemistry of hepatotoxicity. *J Clin Toxicol.* S4:001.
- [49] Soma, B.; Resma, S.; Anjan, A.; Sharmistha, B. and Pratip Kumar, B.(2014): Hepatoprotective activity of *Ixoracoccinea* L. in animal models. *Int J Res Ayurved Pharm.* 5(30):339-42.
- [50] Tadeusz, J.; Teresa, J. and Krzysztof, N. (2001): The role of polyprenol in modulation of physical properties of model membranes. *Curr Top Biophys*;25:33-8.
- [51] Taylor, P.C. (2001). Anti-TNF therapy for rheumatoid arthritis and other inflammatory diseases. *Mol. Biotechnol.* 19, 153–168.
- [52] Walter, M. and Gerade, R.W. (1970): Bilirubin direct /total. *Microchem. J.*, 15:231-233.
- [53] Yengkhom, N.S.; Gunindro, N.; Kholi, S. M.; Moirangthem, R. S. and Rajkumari, B.D. (2017): Hepatoprotective effect of aqueous extract of *Melothriaperpusilla* against carbon tetrachloride induced liver injury in albino rats. *Int J Res Med Sci.* 5:806-10.
- [54] Yousef, M. I.(2004): Aluminium-induced changes in hemato-biochemical parameters, lipid peroxidation and enzyme activities of male rabbits: protective role of ascorbic acid. *Toxicology*. 199:47-57.
- [55] Tsukamoto, H. (2005): Adipogenic phenotype of hepatic stellate cells. *Alcohol. Clin. Exp. Res.* 29, 132S–133S.
- [56] Boll, M.; Weber, L.W.D.; Becker, E. and Stampfl, A. (2001): Pathogenesis of Carbon Tetrachloride-Induced Hepatocyte Injury Bioactivation of CCl4 by Cytochrome P450 and Effects on Lipid Homeostasis. *Z. Naturforsch.* 56c, 111-121.
- [57] Al-Seeni, M. N.; Haddad, A.; El Rabey, H.A.; Zamzami, M.A.; Abeer, M. and Alnefayee, A.M. (2016): The hepatoprotective activity of olive oil and *Nigella sativa* oil against CCl4 induced hepatotoxicity in male rats. *BMC Complement Altern Med.* 16: 438.
- [58] Al-Yahya, M.; Ramzi, M.; Mansour, A.; Mohammed, A. and Nawal, A., et al. (2013): Attenuation of CCl4-induced oxidative stress and hepatonephrotoxicity by Saudi Sidr Honey in rats. *Evidence-Based Complementary and Alternative Medicine*.

- [59] Oloyede, O.B. and Sunmonu, T. O. (2009):Potassium bromate content of selected bread samples in Ilorin, Central Nigeria and its effect on some enzymes of rat liver and kidney. *Food ChemToxicol*, 47: 2067-2070.
- [60] Sabiu, S.; Sunmonu, T.O.; Ajani, E.O. andAjiboye, O.T.(2015): Combined administration of silymarin and vitamin C stalls acetaminophen-mediated hepatic oxidative insults in Wistar rats. *Braz J Pharmacog*, 25: 29-34.
- [61] Sabiu, S.; Wudil, A. M. andSunmonu, T.O. (2014): Combined administration of Telfairaoccidentalis and Vernoniaamygdalina leaf powders ameliorates Garlic-induced hepatotoxicity in Wistar rats. *Pharmacologia*, 5: 191-198.
- [62] Tietz, N.W.(1995): Clinical guide to laboratory tests. 3rd edn. W.B. Saunders, Philadelphia, USA.
- [63] Tan, H.; He, Q.; Li, R.; Lei, F. and Lei, X. (2016): Trillin reduces liver chronic inflammation and fibrosis in carbon tetrachloride (ccl4) induced liver injury in mice. *Immunol Invest*. 45: 371-382.
- [64] Ahn, M.; Kim, J.; Bang, H.; , Moon, J. and Kim, G.O. (2016): Hepatoprotective effects of allylisothiocyanate against carbon tetrachloride-induced
- [65] hepatotoxicity in rat. *ChemBiol Interact* 254: 102-108.
- [66] El-Boshy, M.E.; Fatma,A,F.; Engy, R, E.; Ahmad, A, A.; Gaitha, M and Qustya, N. (2017): Attenuation of CCl4 Induced Oxidative Stress, Immunosuppressive, Hepatorenal Damage by Fucoidan in Rats. *ClinToxicol*, 7:3.
- [67] Shim, J.; Kim, M.; Kim, H.;Ahn, J .; Yun, Y. and Song, J. (2010): Protective action of the immunomodulatorginsan against carbon tetrachloride-induced liver injury via control of oxidative stress and the inflammatory response. *Toxicology and Applied Pharmacology* 242 , 318–325.
- [68] Althnaian, T. (2012):Protective Effect of Camel Milk Against Carbon Tetrachloride Hepatotoxicity in Rats. *Global Veterinaria*,9 (5): 564-570.
- [69] Duncan, D. B. (1955): Multiple range and multiple F. test. *Biometrics*, 11,1.
- [70] Snedecor, G.W. and Cochran, W.G. (1982): statistical methods. 7th ed. P.215, the lawa state univ. Press, Ames, Lawa, USA.
- [71] El Miniawy, H.M.F.; Ahmed, K.A.; Tony, M.A; Mansour, S.;Khattab, M.M.S. (2014): Camel milk inhibits murine hepatic carcinogenesis, initiated by diethylnitrosamine and promoted by phenobarbitone. *International Journal of Veterinary Science and Medicine*. 2: 136–141.
- [72] Ibrahim, M.A.;Wani, F.A. and Rahiman, S. (2017): Hepatoprotective effect of olive oil and camel milk on acetaminophen-induced liver toxicity in mice. *Int J Med Sci Public Health* .6 (1-9).
- [73] Ishak, K.;Baptista, A.; Bianchi, L.;Callea, F.; De Groote, J.;Gudat, F.;Denk, H.;Desmet, V.;Korb, G.;MacSween, R.N.M.; Phillips, M.J.;Portmann, B.G.;Poulsen, H.;Scheuer, P.J.; Schmid, M. and HeribertThaler, H. (1995):Histologic grading and staging of chronic hepatitis. *J Hepatol*, 24:289-293.
- [74] Shackelford, C., long, G., Wolf,J., Okerberg, C., and Herbert. R. (2002): Qualitative and quantitative analysis of nonneoplastic lesionsIn toxicology studies. *Toxicologic pathology*,30:(1) 93–96.
- [75] Grassi, J., Roberge, C.J and Frobert, Y. (1991): Determination of IL-1 α , 1L-1 β and IL-2 in biological media using specific enzyme immunometric assay. *Immunol. Res.*, 119:125-145.
- [76] NRC (National Research Council), (1977): Nutrient Requirements of Domestic Animals, National Academy of Science, Washington DC, USA.
- [77] Haversen, L.;Ohlsson, B.G.; Hahn-Zoric, M.; Hanson, L.A.;Mattsby-Baltzer, I. (2002):
- [78] Lactoferrin down-regulates the LPS-induced cytokine production in monocytic
- [79] cells via NF-kappa B. *Cell. Immunol*. 220, 83–95.
- [80] Desai S. N.; Patel, D.K.; Devlar, R.V.; Patel, P.V.; and Amachandran, A.V. (2012): Hepatoprotectivepotentialof polyphenol rich extract Of Murrayakoenigii.: an in vivo study. *Food ChemToxicol*. 50:310-314.
- [81] ALHAIDER,A.A., . ABDEL GADER, A.M., ALMESHAAL, N. and SARASWATI, S. (2013): Camel milk inhibits inflammatory angiogenesis via downregulation of proangiogenic and proinflammatory cytokines in mice. *APMIS*, 122: 599–607.