

# Research Journal of Pharmaceutical, Biological and Chemical Sciences

## A Comprehensive Review of Benzo Alpha Pyrene (B[A]P) Toxicology.

Sandeep Agrawal<sup>1</sup>, Amar Preet Kaur<sup>1\*</sup>, and Kanchan Taneja<sup>2</sup>.

<sup>1</sup>Department of Biochemistry, All India Institute of Medical Sciences (AIIMS), Ansari Nagar, New Delhi-110029; India

<sup>2</sup>Department of Biochemistry, Chacha Nehru Bal Chikitsalaya, Geeta colony, New Delhi-110031, India.

### ABSTRACT

Benzo Alpha Pyrene (B[a]P) is a prototype Polycyclic Aromatic Hydrocarbon (PAH). It is a well known environmental pollutant and a procarcinogen. It is present ubiquitously in the environment and exposure to B[a]P occurs through air, food, water, skin contact etc. It is metabolized in the body by phase I and phase II reactions and the primary metabolites are non-toxic and easily excretable. A minor fraction of B[a]P is converted to active metabolites which are carcinogenic and adversely affect other systems of the body. This review covers all aspects of B[a]P including sources, exposure, metabolism and toxicity profile.

**Keywords:** Benzo alpha pyrene (B[a]P), Polycyclic Aromatic Hydrocarbon (PAH), carcinogen, Cytochrome P450

*\*Corresponding author*

## INTRODUCTION

Polycyclic aromatic hydrocarbons (PAH) are produced by incomplete combustion of organic compounds. A 5-ring structure compound Benzo[a]Pyrene (B[a]P) is a prototype of PAH. B[a]P is an environmental pollutant, a carcinogen and, is ubiquitously present in our surroundings i.e. air, water, soil, charcoal grilled food items, certain pharmaceutical products and tobacco smoke [1,2]. Sources of B[a]P in ambient air are industrial emissions, motor vehicle exhaust, tobacco smoke, cooking and residential and commercial heating of organic fuel etc. Concentration of B[a]P is found to be more than three times higher in side stream cigarette smoke than mainstream smoke.

Sources of B[a]P in the diet are barbecued/grilled/broiled and smoke cured meats, roasted and baked foods and vegetables grown in contaminated soil [2] (Figure 1). B[a]P is also present in soil. The levels of B[a]P in the soil vary depending on proximity to roads, combustion sources, use of sewage or sludge derived amendments on agricultural lands etc. Exposure can also occur via the use of dermally applied pharmaceutical products which contain coal tars (including formulations used to treat eczema and psoriasis) [2]. Use of contaminated water (by petroleum spills, road run off, industrial wastewater) also increases the risk of exposure to B[a]P [3, 4, 5].

Occupational exposure occurs through inhalation and skin contact. B[a]P exposure has been reported in occupations related to coal liquefaction, coal gasification, coke production and coke ovens, coal-tar distillation, roofing and paving (involving coal-tar pitch), aluminium production (including anode manufacture), carbon-electrode manufacture, chimney sweeping, and power plants [2].

## METABOLISM

B[a]P enters the body mainly through inhalation and ingestion. It is then transported to various organs through blood and lymph [6]. B[a]P is metabolized by both phase-I and phase-II enzymes to form epoxides, dihydrodiols, phenols, and quinones and their polar conjugates with glutathione, sulphate and glucuronide [7].

After ingestion, absorption, and transport, the initial oxygenation of B[a]P is catalysed by the microsomal mixed-function oxidases (MFOs), which contains multiple forms of cytochrome P-450 [8, 9]. The major cytochrome P450s involved in the B[a]P metabolism are CYP1A1, CYP1A2 and CYP1B1 [10, 11]. Cytochrome P450s are inducible by B[a]P and other PAHs through binding to the aryl hydrocarbon-receptor (AhR) nuclear complex, resulting in changes in gene transcription of CYPs and phase-II enzymes.

Primary metabolites of B[a]P metabolism (Figure 2):

1. Three epoxides: 4,5-epoxide, 7,8-epoxide and 9,10-epoxide [9, 12, 13].
2. Three dihydrodiols: (-)-trans-4,5-diol, (-)-trans-7,8-diol and (-)-trans-9,10-diol [14, 15, 16, 17].
3. Five phenols: 1-OH, 3-OH, 6-OH, 7-OH and 9-OH. These phenols can be converted to quinones [18, 19, 20, 21, 22].

CYP450 acts on B[a]P leading to formation of epoxides at the 4,5-, 7,8-, and 9, 10-positions. Further epoxide hydratase acts on the epoxide intermediates [23, 24] to form corresponding dihydrodiols. Few epoxide intermediates get converted to phenols. The major phenol metabolite of B[a]P is 3-hydroxybenzo[a]pyrene (3-OHBP) [25, 26] formed in presence of an enzyme aryl hydrocarbon hydroxylase. The 7- and 9-phenols are rearrangement products of the unstable 7,8- and 9,10-epoxide intermediates [23, 27, 28]. Other phenols may be formed either by direct hydroxylation or rearrangement of the unstable epoxide intermediates.

The primary epoxides can be conjugated to glutathione S-conjugates [29] and the phenols and diols can be conjugated to either sulphate [30] or glucuronide [31] to form water-soluble compounds. The formation of the water-soluble glutathione, glucuronide, and sulphate conjugates is catalysed by glutathione S-epoxide transferase, UDPglucuronate transferase, and sulfotransferase, respectively. The majority of B[a]P is converted to these water-soluble, easily excretable non-toxic metabolites.

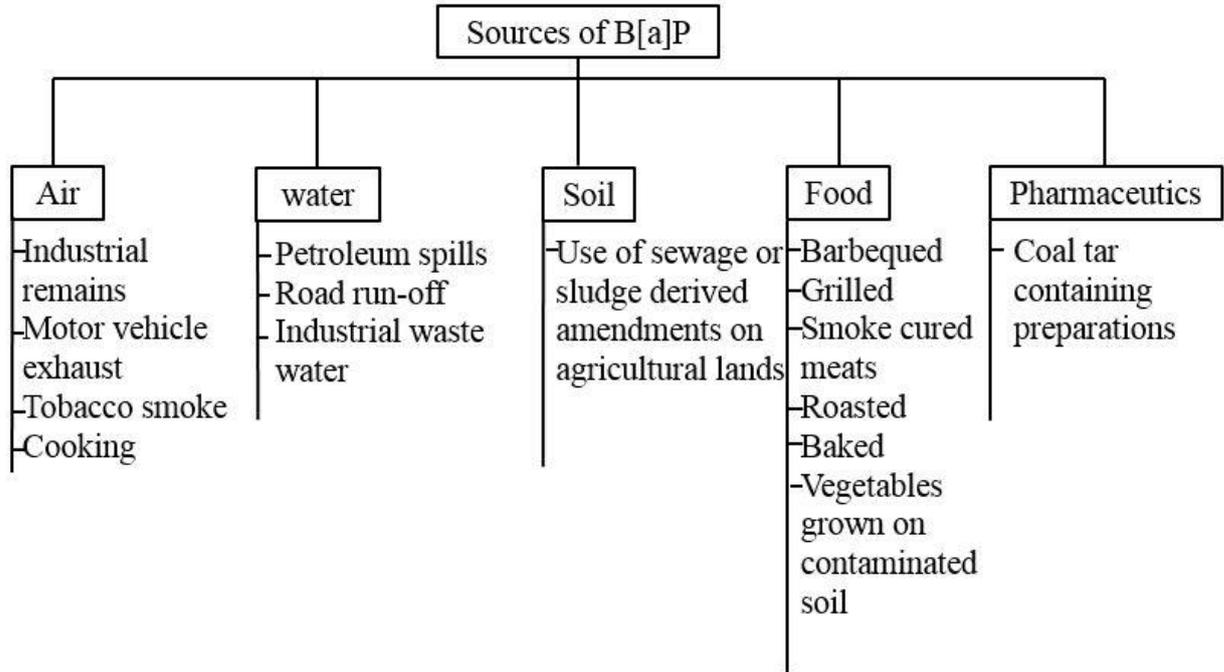


Figure 1: Sources of B[a]P exposure

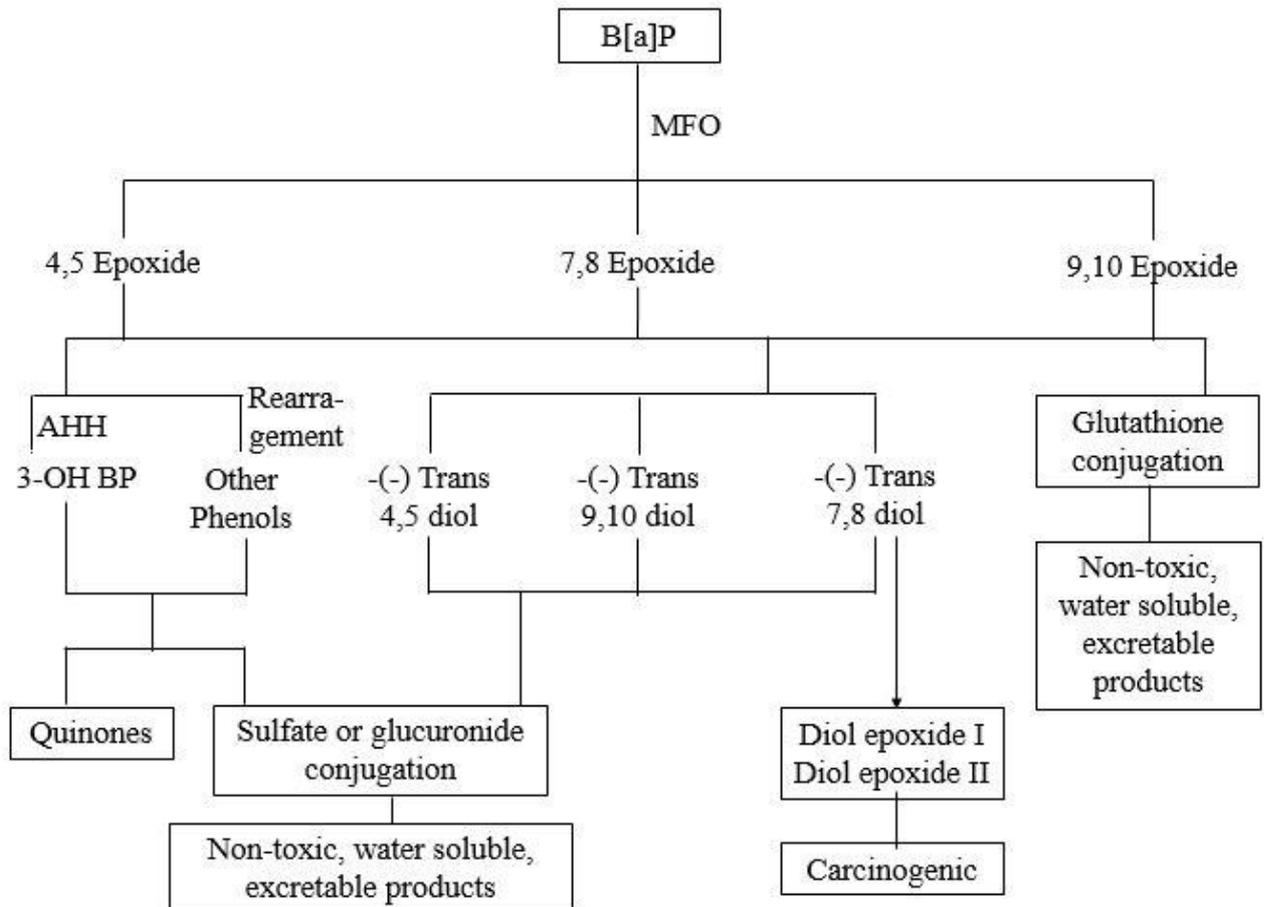


Figure 2: Overview of B[a]P metabolism

A small fraction of B[a]P is converted to active metabolites which are genotoxic and carcinogenic [8] by further metabolism or “recycling” of the phenol and dihydrodiol metabolites through the MFO system. Thus, the (-)-trans-7,8-diol is converted to two stereoisomers of 7,8-diol-9,10-epoxides: diol epoxide I and diol epoxide II [32, 33] by the MFOs. There are four possible stereoisomeric diol epoxides derived from the trans-7,8-diol. Each diol epoxide is further hydrolysed to two tetrols and reduced to a triol.

The diol epoxides are highly carcinogenic metabolites of B[a]P. The amount of recycling of B[a]P metabolites by the MFOs to further oxidized products depends on the experimental conditions. Factors that control recycling depend on the availability of phenol or diol available to the MFO. Less recycling occurs when large amounts of B[a]P substrates are present and a large amount of recycling is observed when very small amounts of B[a]P are used as substrate [34].

Second factor governing the frequency of recycling is the presence of competing enzyme systems with the appropriate cofactors. High levels of conjugating enzymes compete with the MFOs for the oxygenated metabolites of B[a]P, thus decreasing the probability of oxygenated metabolites for recycling.

### EFFECTS OF B[a]P METABOLISM

**Binding of B[a]P Metabolites with Macromolecules:** The metabolic intermediates of B[a]P metabolism bind covalently to nucleic acids [35], resulting in cytotoxicity, mutagenicity [36], cell transformation in vitro and cancer induction in experimental animals.

B[a]P diol epoxides I and II form covalent adducts primarily with guanosine residues of RNA and DNA. Up to a limited extent, they can also form covalent adducts with adenosine and cytidine [37]. The carbon at the 10<sup>th</sup> position of the B[a]P diol epoxide is linked to the N-2-amino group of guanines.

Studies done to decipher the carcinogen nature of B[a]P indicate that out of the four stereoisomers of the 7,8-diol-9,10 epoxide, the (+) diol epoxide I has higher tumorigenicity as compared to other 3 stereoisomers [38]. Also, the tumor initiating activity of (+) diol epoxide I was much higher as compared to other stereoisomers. Diol epoxides are the predominant metabolites as carcinogen and tumor initiating forms but other routes of activation of B[a]P to other metabolites may also contribute to carcinogenesis.

B[a]P diol epoxides also bind covalently to cellular proteins. Weinstein et al., (1976) demonstrated the interaction between B[a]P diol epoxides and protein components of chromatin.

**Cancer:** Even after extensive literature review, we could not find any epidemiological data of B[a]P carcinogenesis in humans. B[a]P is known to cause tumor in experimental animals following exposure through many different routes which are summarized below.

1. **Oral administration:** There was an increase in incidence of tumors in various organs like tongue, liver, lung, forestomach, oesophagus, lymphoid tissue and haematopoietic tissues after oral administration of B[a]P either by gavage or in the diet of mice [39-47].
2. **Inhalation:** B[a]P induced increase in incidence of papilloma and squamous cell carcinoma in both upper respiratory tract and upper digestive tract in male hamsters [48].
3. **Skin application:** Benign (Squamous cell papillomas and Keratoacanthomas) and malignant (Squamous cell carcinoma) tumors were observed in different strains of mice when B[a]P was applied directly on skin [49-58].
4. **Subcutaneous (s.c.) injection:** Fibrosarcomas (malignant tumors) developed at the injection site in mice [59-62].
5. **Intraperitoneal (i.p.) injection:** Intraperitoneal injection of B[a]P in newborn and adult mice led to an increased incidence of liver (adenomas and carcinomas), lung (adenomas and carcinomas) and lymphoreticular tumors [63-73].
6. **Intrapulmonary injection:** After injection of B[a]P in the lung of rats, there was an increase in the incidence of malignant lung tumors (mainly squamous cell carcinomas) [74-77].
7. **Intratracheal administration:** Intratracheal administration of B[a]P resulted in benign and malignant respiratory tumors in mice, rats and in hamsters [78-80].

8. **Buccal pouch application:** A higher incidence of forestomach papillomas was observed in male hamsters after repeated application of B[a]P to the buccal pouch mucosa[81].
9. **Intramammary administration:** Benign and malignant mammary gland tumors were observed after intramammary injection of B[a]P in rats[51, 82, 83].
10. **Intracolonic instillation:** Intracolonic instillation of B[a]P induced lymphomas in forestomach and various other organs in mice[84, 85].
11. **Intravaginal application:** In mice, intravaginal application of B[a]P produced invasive cervical carcinoma[86].
12. **Intrafetal injection:** One study conducted by Rossi et al.,[87] showed that intrafetal injection of B[a]P produced lung adenomas in male and female Swiss mice.

**Epigenetic effects:** The metabolites of B[a]P have been found to increase cell proliferation and increased expression of the *Cdc25B* gene (cell-division cycle25B) and reduced phosphorylation of Cdk1 (cyclin-dependent kinase1) in different human cell lines[88]. Exposure to B[a]P and its metabolites leads to alteration in DNA methylation. After treatment of immortalized bronchial epithelial cells with anti-B[a]P-7,8-diol-9,10-epoxide, the concentration of cytosine-DNA methyltransferase-1 was increased and was associated with hypermethylation of the promoters of 5–10 genes, including members of the cadherin gene-family[89].

**Apoptosis:** B[a]P and its metabolites also regulate apoptosis. B[a]P was found to induce apoptosis via the JNK1/FasL (c-Jun N-terminal kinase 1/FasLigand) and JNK1/p53 signalling pathways in human MRC-5 lung fibroblasts [90]. Apoptosis induced by anti-B[a]P-7,8-diol-9,10-epoxide in H460 human lung-cancer cells was associated with induction of Bak (BCL2-antagonist) and with activation of caspase [91].

**Effects on other systems:** Following oral exposure, animal studies have demonstrated developmental neurotoxicity, reproductive toxicity and immunotoxicity. Gestational exposure to mice and rats lead to neurobehavioral changes and cardiovascular effects. Oral exposure in adult animals lead to various reproductive and immune system dysfunction including decrease in sperm count, ovary weight, follicle numbers and decrease in thymus weight, B cell numbers and immunoglobulin concentration. B[a]P suppresses immunity by modulating p53-dependent signalling pathways in lymphocytes. B[a]P has been shown to induce immunosuppression in adult mice by altering the cell-mediated responses [92]. Immune development in offspring was also altered following *in utero* exposure to B[a]P[93].

Following inhalational exposure, developmental and reproductive toxicity was observed in rats which included decreased fetal survival, nervous system defects in offspring and decreased testes weight and sperm counts in adult animals[94].

## DISCUSSION

B[a]P, a well characterized procarcinogen, is a member of the PAH family. It is ubiquitously present in the environment and enters the body mainly by inhalation and ingestion. Once inside the cells, it is metabolized by P450-dependent monooxygenase system to various epoxides, phenols and dihydro diols. These metabolites are either converted to water soluble excretory products or recycled to reactive and toxic metabolites depending upon the availability of substrates and enzymes of the two pathways.

The adverse effects of B[a]P includes carcinogenesis, teratogenicity, neurotoxicity, reproductive toxicity, immunosuppression and developmental toxicity in experimental animals. It also affects lipid metabolism, apoptosis and induces epigenetic modifications. B[a]P is categorized as a human Group 1 carcinogen by the International Agency for Research on Cancer [95]. The location of tumors depends on the route of exposure. Inhalation of B[a]P often induces lung cancer, while oral administration leads to tumors in various organs/tissues, including the gastro-intestinal tract, liver, lungs, and mammary glands [96].

Most of the information on B[a]P toxicity has been obtained from animal studies and this data cannot be extrapolated for human trials because of species difference. Also the higher toxicological doses which are used in animal testing might already be present at relevant concentrations in environment. These disadvantages limit the extrapolation of data from animal studies to humans.

Therefore, the US National Research Council proposed the Toxicity Testing in the 21st Century (TT21C) and encouraged the use of *in vitro* toxicity pathway-based approaches using cell lines (97), as compared to high-dose studies in laboratory animals. The toxicity pathways, or adverse outcome pathways (AOP) are evaluated by *in vitro* assays of innate cellular signalling pathways and are severely affected if disturbed[97]. However, the AOP/TT21C strategy in B[a]P toxicity testing is still under investigation.

Based on the best available literature, it is concluded that B[a]P contributes to various deleterious effects including carcinogenesis and toxicity of various organ systems. The strong and extensive experimental evidence for the carcinogenicity of B[a]P in many animal species and human cell lines support the overall classification of B[a]P as a human carcinogen (Group 1).

#### REFERENCES

- [1] Phillips DH (1983). Fifty years of benzo(a)pyrene. *Nature* 303:468-472.
- [2] IARC 2012 Monographs. Chemical Agents and Related Occupations. IARC monographs on the evaluation of carcinogenic risks to humans. 100F:111-144.
- [3] Ngabe B, Bidleman TF, Scott GI (2000). Polycyclic aromatic hydrocarbons in storm runoff from urban and coastal South Carolina. *Sci Total Environ*, 255(1-3):1-9.
- [4] Vogelsang C, Grung M, Jantsch TG, Tollefsen KE, Liltved H (2006). Occurrence and removal of selected organic micropollutants at mechanical, chemical and advanced wastewater treatment plants in Norway. *Water Res*, 40(19):3559-3570
- [5] Vidal M, Domínguez J, Luís A (2011). Spatial and temporal patterns of polycyclic aromatic hydrocarbons (PAHs) in eggs of a coastal bird from northwestern Iberia after a major oil spill. *Sci Total Environ*, 409(13):2668-2673.
- [6] Van de Wiele T, Vanhaecke L, Boeckeaert C, Peru K, Headley J, Verstraete W, et al (2005). Human colon microbiota transform polycyclic aromatic hydrocarbons to estrogenic metabolites. *Environ Health Perspect* 113:6-10.
- [7] Osborne MR, Crosby NT (1987). *Benzopyrenes*. Cambridge Monographs on Cancer Research. Cambridge, England: Cambridge University Press.
- [8] Schellenberger MT, Grova N, Willième S, Farinelle S, Prodhomme EJ, Muller CP (2009). Modulation of benzo[a]pyrene induced immunotoxicity in mice actively immunized with a B[a]P-diphtheria toxoid conjugate. [Toxicology and Applied Pharmacology](#). 240(1):37-45.
- [9] Selkirk, J. K., R. G. Croy, and H. V. Gelboin (1975). Isolation and characterization of benzo[u]pyrene. *Arch. Biochem. Biophys.* 168: 322-326.
- [10] Eling T, Curtis J, Battista J, Marnett LJ (1986). Oxidation of (+)-7,8-dihydroxy-7,8-dihydrobenzo[a]pyrene by mouse keratinocytes: evidence for peroxy radical- and monooxygenase-dependent metabolism. *Carcinogenesis*, 7: 1957-1963.
- [11] Shimada T (2006). Xenobiotic-metabolizing enzymes involved in activation and detoxification of carcinogenic polycyclic aromatic hydrocarbons. *Drug Metab Pharmacokinet*, 21: 257-276.
- [12] Sims, P., and P. L (1974). Grover. Epoxides in polycyclic aromatic hydrocarbon metabolism and carcinogenesis. *Adv. Cancer Res.* 20: 165-274.
- [13] Yang, S. K., P. P. Roller, P. P. FU, R. G. Harvey, and H. V. Gelboin (1977). Evidence for a 2,3-epoxide as an intermediate in the microsomal metabolism of benzo[a]pyrene to 3-hydroxybenzo[a]pyrene. *Biochem. Biophys. Res. Commun.* 77: 1176-1182.
- [14] Booth, J., and P. Sims (1976). Different pathways involved in the metabolism of the 7,8- and 9,10-dihydrodiols of benzo[a]pyrene. *Biochem. Pharmacol.* 25: 979-980.
- [15] Borgen, A., H. Davey, N. Castagnoli, T. T. Crocker, R. E. Rasmussen, and I. Y. Wang (1976). Metabolic conversion of benzo[u]pyrene by Syrian hamster liver microsomes and binding of metabolites to deoxyribonucleic acid. *J. Med. Chem.* 16: 502-506.
- [16] Pezzuto, J. M., C. S. Yang, S. K. Yang, D. W. McCourt, and H. V. Gelboin (1978). Metabolism of benzo[u]pyrene and (-)-trans-7,8-dihydroxy-7,8-dihydrobenzo[u]pyrene by rat liver nuclei and microsomes. *Cancer Res.* 38: 1241-1245.
- [17] Selkirk, J. K., R. G. Croy, and H. V. Gelboin (1974). Benzo[u]pyrene metabolites: efficient and rapid separation by high-pressure liquid chromatography. *Science* 184:169-171.
- [18] Conney, A. H., E. C. Miller, and J. A. Miller (1957). substrate-induced synthesis and other properties of benzopyrene hydroxylase in rat liver. *J. Biol. Chem.* 228:753-766.

- [19] Engel, and H. V. gelboin (1976). Separation of ten benzo[u]pyrene phenols by recycle high pressure liquid chromatography and identification of four phenols as metabolites. *Biochem. Phurmucol.* 25: 227-230.
- [20] Falk, H. L., P. Kotin, S. S. Lee, and A. Nathan (1962). Intermediary metabolism of benzo(u)pyrene in the rat. *J.Nutl. Cancer Inst.* 28: 699-724.
- [21] freudenthal, R. I., A. P. Leber, D. Emmerling, and P. Clarke (1975). The use of high pressure liquid chromatography to study chemically induced alterations in the pattern of benzo(u)pyrene metabolism. *Chem. Biol. Interact.* 11: 449-458.
- [22] Gelboin (1976). Identification of mutagenic metabolites of benzo[a]pyrene in mammalian cells. *Proc. Natl. Acad.Sci. USA* 73: 607-611.
- [23] Holder, G., H. Yagi, P. Dansette, D. M. Jerina, W. Levin, A. Y. H. LU, and A. H. Conney (1974). Effects of inducers and epoxide hydrase on the metabolism of benzo[u]pyrene by liver microsomes in a reconstituted system: analysis by high pressure liquid chromatography. *Proc. Nutl. Acad. Sci. USA* 71:4356-4360.
- [24] Yang, S. K., P. P. Roller, and H. V. Gelboin (1977). Enzymatic mechanism of benzo[a]pyrene conversion to diols and phenols and an improved high-pressure liquid chromatographic separation of benzo[a]pyrene derivatives. *Biochemistry* 16: 3680-3687.
- [25] Berenblum, I., D. Crowfoot, B. R. Holiday, and R. Schoental (1943). The metabolism of 3,4-benzpyrene in mice and rats. II. The identification of the isolated products as 8-hydroxy-3,4-benzpyrene and 3,4-benzpyrene- 5,&quinone. *Cancer Res.* 3: 151-158.
- [26] Berenblum, I., and R. Schoental (1955). Metabolism of 3,4-benzopyrene. *Science* 122: 470.
- [27] Sims, P., and P. L. Grover (1974). Epoxides in polycyclic aromatic hydrocarbon metabolism and carcinogenesis. *Adv. Cancer Res.* 20: 165-274.
- [28] Waterfall, J. F., and P. Sims (1972). Epoxy derivatives of aromatic polycyclic hydrocarbons. The preparation and metabolism of epoxides related to benzo[u]pyrene and to 7,8- and 9,10-dihydrobenzo[u]pyrene. *Biochem. J.* 128:265-277.
- [29] Nemoto, N., and H. V. Gelboin (1975). Assay and properties of glutathione-S-benzo[u]pyrene-4,5-oxide transferase. *Arch. Biochem. Biophys.* 170: 739-742.
- [30] Nemoto, N., and S. Takayama (1977). Enzymatic formation and properties of a conjugate of sulfate with 3-hydroxybenzo[a]pyrene. *Biochem. Pharmacoz.* 26: 679-684.
- [31] Nemoto, N., and H. V. Gelboin (1976). Enzymatic conjugation of benzo[a]pyrene oxide phenols and dihydrodiols with UDP-glucuronic acid. *Biochem. Pharmacol.* 25:1221- 1226.
- [32] Huberman, E., L. Sachs, S. K. Yang, and H. V. Gelboin (1976). Identification of mutagenic metabolites of benzo[a]pyrene in mammalian cells. *Proc. Natl. Acad.Sci. USA* 73: 607-611.
- [33] Yang, S. K., D. W. Mccourt, J. C. Leutz, and H. V. Gelboin (1977). Benzo[a]pyrene diol-epoxides: mechanism of enzymatic formation and optically active intermediates. *Science* 196: 1199-1201.
- [34] Holder, G. M., H. Yagi, D. M. Jerina, W. Levin, A. Y. H. LU, and A. H. Conney (1975). Metabolism of benzo[u]pyrene: effect of substrate concentration and 3-methylcholanthrene pretreatment of hepatic metabolism by microsomes from rat and mice. *Arch. Biochem.Biophys.* 170: 557-566.
- [35] Alexandrov, K., and M. H. Thompson (1977). Influence of inducers and inhibitors of mixed-function oxidases on benzo(a)pyrene binding to the DNA of rat liver nuclei. *Cancer Res.* 37: 1443-1449.
- [36] Ames, B. N., W. C. Durston, E. Yamasaki, and F. D. LEE (1973). Carcinogens are mutagens: a simple test system combining liver homogenates for activation and bacteria for detection. *Proc. Natl. Acad. Sci. USA* 70: 2281-2285.
- [37] Weinstein, I. B., A. M. Jeffrey, S. Leffler, P. Pulkrabek, H. Yamasaki, and D. Grunberger(1978). Interactions between polycyclic aromatic hydrocarbons and cellular macromolecules. In: *Polycyclic Hydrocarbons and Cancer*, edited by H. V. Gelboin and P. O. P. Ts'o. New York: Academic. 2:4-36.
- [38] Buening, M. K., P. G. Wislocki, W. Levin, H. Yagi, D. R. Thakker, H. AKAGI, M. Koreeda, D. M. Jerina, and A. H. Conney (1978). Tumorigenicity of the optical enantiomers of the diastereomeric benzo(u)-pyrene 7,8diol, 9,10-epoxides in newborn mice: exceptional activity of (+)-7p, Ba-dihydroxyl-9a, lOa-epoxy-7,8,9, lotetrahydrobenzo (u)pyrene. *Proc. N&Z. Acad. Sci. USA* 75: 5358-5361.
- [39] Sparnins VL, Mott AW, Barany G, Wattenberg LW (1986). Effects of allyl methyl trisulfide on glutathione S-transferase activity and BP-induced neoplasia in the mouse. *Nutrition and Cancer*, 8: 211–215.
- [40] Estensen RD & Wattenberg LW (1993). Studies of chemopreventive effects of myo-inositol on benzo[a]pyreneinduced neoplasia of the lung and forestomach of female a. A/J mice. *Carcinogenesis*, 14: 1975–1977.

- [41] Weyand EH, Chen Y-C, Wu Y *et al.* (1995). Differences in the tumorigenic activity of a pure hydrocarbon and a complex mixture following ingestion: benzo[a]pyrene vs manufactured gas plant residue. *Chemical Research in Toxicology*, 8: 949–954.
- [42] Kroese ED, Dortant PM, van Steeg H *et al.* (1997). Use of E  $\mu$ -PIM-1 transgenic mice short-term in vivo carcinogenicity testing: lymphoma induction by benzo[a]pyrene, but not by TPA. *Carcinogenesis*, 18: 975–980.
- [43] Culp SJ, Gaylor DW, Sheldon WG *et al.* (1998). A comparison of the tumors induced by coal tar and benzo[a]pyrene in a 2-year bioassay. *Carcinogenesis*, 19: 117–124.
- [44] Hakura A, Tsutsui Y, Sonoda J *et al.* (1998). Comparison between in vivo mutagenicity and carcinogenicity in multiple organs by benzo[a]pyrene in the lacZ transgenic mouse (Muta Mouse). *Mutat Res*, 398: 123–130.
- [45] Badary OA, Al-Shabanah OA, Nagi MN *et al.* (1999). Inhibition of benzo[a]pyrene-induced forestomach carcinogenesis in mice by thymoquinone. *European Journal of Cancer Prevention*, 8:435–440.
- [46] Wijnhoven SWP, Kool HJM, van Oostrom CTM *et al.* (2000). The relationship between benzo[a]pyrene-induced mutagenesis and carcinogenesis in repair deficient Cockayne syndrome group B mice. *Cancer Res*, 60: 5681–5687.
- [47] Estensen RD, Jordan MM, Wiedmann TS *et al.* (2004). Effect of chemopreventive agents on separate stages of progression of benzo[alpha]pyrene induced lung tumors in A/J mice. *Carcinogenesis*, 25: 197–201.
- [48] Thyssen J, Althoff J, Kimmerle G, Mohr U (1981). Inhalation studies with benzo[a]pyrene in Syrian golden hamsters. *J Natl Cancer Inst*, 66: 575–577.
- [49] Van Duuren BL, Katz C, Goldschmidt BM (1973). Cocarcinogenic agents in tobacco carcinogenesis. *J Natl Cancer Inst*, 51: 703–705.
- [50] Cavaliere E, Mailander P, Pelfrene A (1977). Carcinogenic activity of anthanthrene on mouse skin. *Zeitschrift fur Krebsforschung*, 89: 113–118.
- [51] Cavaliere E, Rogan E, Cremonesi P *et al.* (1988a). Tumorigenicity of 6-halogenated derivatives of benzo[a] pyrene in mouse skin and rat mammary gland. *Journal of Cancer Research and Clinical Oncology*, 114: 10–15.
- [52] Levin W, Wood AW, Wislocki PG *et al.* (1977). Carcinogenicity of benzo ring derivatives of benzo[a]pyrene on mouse skin. *Cancer Research*, 37: 3357–3361.
- [53] Habs M, Jahn SAA, Schmahl D (1984). Carcinogenic activity of condensate from colocynth seeds (*Citrullus colocynthis*) after chronic epicutaneous administration to mice. *J Cancer Res Clin Oncol*, 108: 154–156.
- [54] Habs M, Schmahl D, Misfeld J (1980). Local carcinogenicity of some environmentally relevant polycyclic aromatic hydrocarbons after lifelong topical application to mouse skin. *Arch Geschwulstforsch*, 50: 266–274.
- [55] Warshawsky D & Barkley W (1987). Comparative carcinogenic potencies of 7H-dibenzo[c,g]carbazole, dibenz[a,j]acridine and benzo[a]pyrene in mouse skin. *Cancer Letters*, 37: 337–344.
- [56] Warshawsky D, Barkley W, Bingham E (1993). Factors affecting carcinogenic potential of mixtures. *Fundamental and Applied Toxicology*, 20: 376–382.
- [57] Albert RE, Miller ML, Cody T *et al.* (1991). Benzo[a]pyrene-induced skin damage and tumor promotion in the mouse. *Carcinogenesis*, 12: 1273–1280.
- [58] Andrews J, Halliday GM, Muller HK (1991). A role for prostaglandins in the suppression of cutaneous cellular immunity and tumour development in benzo[a]pyrenebut not dimethylbenz(a)anthracene-treated mice. *Clin Exp Immunol*, 85: 9–13.
- [59] Kouri RE, Wood AW, Levin W *et al.* (1980). Carcinogenicity of benzo[a]pyrene and thirteen of its derivatives in C3H/fCum mice. *J Natl Cancer Inst*, 64: 617–623.
- [60] Rippe RM, Pott D (1989). Kanzerogenitätsuntersuchungen von Nitro-PAH (Nitroarenen) im Hinblick auf ihre Bedeutung für die krebserzeugende Wirkung von Dieselmotorabgas. In: Gesellschaft zur Förderung der Lufthygiene und Silikoseforschung. Dusseldorf: Stefan W. Albers, pp. 65–89.
- [61] Pott F, Brockhaus A, Huth F (1973a). [Tests on the production of tumours in animal experiments with polycyclic aromatic hydrocarbons.] [in German]. *Zbl. Bakt. Hyg. Abt. Orig. B*, 157: 34–43.
- [62] Pott F, Tomingas R, Reiffer FJ (1973b). Experimental studies on the carcinogenicity and the retention of benzo[a]pyrene in application region after intratracheal and subcutaneous injection. *Zbl. Bakt. Hyg. I. Abt. Orig. B*, 158: 97–108.

- [63] Vesselinovitch SD, Kyriazis AP, Mihailovich N, Rao KVN (1975a). Factors influencing augmentation and/or acceleration of lymphoreticular tumors in mice by benzo[a]pyrene treatment. *Cancer Res*, 35: 1963–1969.
- [64] Vesselinovitch SD, Kyriazis AP, Mihailovich N, Rao KVN(1975b). Conditions modifying development of tumors in mice at various sites by benzo[a]pyrene. *Cancer Res*, 35: 2948–2953.
- [65] Wislocki PG, Bagan ES, Lu AY *et al.* (1986). Tumorigenicity of nitrated derivatives of pyrene, benz[a]anthracene, chrysene and benzo[a]pyrene in the newborn mouse assay. *Carcinogenesis*, 7: 1317–1322.
- [66] Lavoie EJ, Braley J, Rice JE, Rivenson A (1987). Tumorigenic activity of non-alternant polynuclear aromatic hydrocarbons in newborn mice. *Cancer Letters*, 34: 15-20.
- [67] Busby WF Jr, Stevens EK, Martin CN *et al.* (1989). Comparative lung tumorigenicity of parent and mononitro-polynuclear aromatic hydrocarbons in the BLU:Hanewborn mouse assay. *Toxicology and Applied Pharmacology*, 99: 555–563.
- [68] Mass MJ, Jeffers AJ, Ross JA *et al.* (1993). Ki-ras oncogene mutations in tumors and DNA adducts formed by benz[j]aceanthrylene and benzo[a]pyrene in the lungs of strain A/J mice. *Molecular Carcinogenesis*, 8:186–192.
- [69] Nesnow S, Ross JA, Stoner GD, Mass MJ (1995). Mechanistic linkage between DNA adducts, mutations in oncogenes and tumorigenesis of carcinogenic environmental polycyclic aromatic hydrocarbons in strain A/J mice. *Toxicology*, 105: 403–413.
- [70] Ross JA, Nelson GB, Wilson KH *et al.* (1995). Adenomas induced by polycyclic aromatic hydrocarbons in strain A/J mouse lung correlate with time-integrated DNA adduct levels. *Cancer Res*, 55: 1039–1044.
- [71] Weyand EH, Chen Y-C, Wu Y *et al.* (1995). Differences in the tumorigenic activity of a pure hydrocarbon and a complex mixture following ingestion: benzo[a]pyrene vs manufactured gas plant residue. *Chemical Research in Toxicology*, 8: 949–954.
- [72] Rodriguez LV, Dunsford HA, Steinberg M *et al.* (1997). Carcinogenicity of benzo[a]pyrene and manufactured gas plant residues in infant mice. *Carcinogenesis*, 18:127–135.
- [73] Von Tungeln LS, Xia Q, Herrero-Saenz D *et al.* (1999). Tumorigenicity of nitropolycyclic aromatic hydrocarbons in the neonatal B6C3F1 mouse bioassay and characterization of ras mutations in liver tumors from treated mice. *Cancer Letters*, 146: 1–7.
- [74] Deutsch-Wenzel RP, Brune H, Grimmer G *et al.* (1983). Experimental studies in rat lungs on the carcinogenicity and dose-response relationships of eight frequently occurring environmental polycyclic aromatic hydrocarbons. *J Natl Cancer Inst*, 71: 539–544.
- [75] Iwagawa M, Maeda T, Izumi K *et al.* (1989). Comparative dose-response study on the pulmonary carcinogenicity of 1,6-dinitropyrene and benzo[a]pyrene in F344 rats. *Carcinogenesis*, 10: 1285–1290.
- [76] Wenzel-Hartung R, Brune H, Grimmer G *et al.* (1990). Evaluation of the carcinogenic potency of 4 environmental polycyclic aromatic compounds following intrapulmonary application in rats. *Exp Pathol*, 40:221–227.
- [77] Horikawa K, Sera N, Otofujii T *et al.* (1991). Pulmonary carcinogenicity of 3,9- and 3,7-dinitrofluoranthene, 3-nitrofluoranthene and benzo[a]pyrene in F344 rats. *Carcinogenesis*, 12: 1003–1007.
- [78] Heinrich U, Pott F, Mohr U *et al.* (1986a). Lung tumours in rats and mice after inhalation of PAH-rich emissions. *Exp Pathol*, 29: 29–34.
- [79] Pott F, Ziem U, Reiffer F-J *et al.* (1987). Carcinogenicity studies on fibres, metal compounds, and some other dusts in rats. *Exp Pathol*, 32: 129–152.
- [80] Steinhoff D, Mohr U, Hahnemann S (1991). Carcinogenesis studies with iron oxides. *Exp Pathol*, 43: 189–194.
- [81] Solt DB, Polverini PJ, Calderon L (1987). Carcinogenic response of hamster buccal pouch epithelium to 4 polycyclic aromatic hydrocarbons. *Journal of Oral Pathology*, 16: 294–302.
- [82] Cavalieri E, Rogan E, Sinha D (1988b). Carcinogenicity of aromatic hydrocarbons directly applied to rat mammary gland. *J. Cancer clin. Oncol*, 114: 3–9.
- [83] Cavalieri EL, Higginbotham S, RamaKrishna NVS *et al.* (1991). Comparative dose-response tumorigenicity studies of dibenzo[a,l]pyrene versus 7,12-dimethylbenz[a]anthracene, benzo[a]pyrene and two dibenzo[a,l]pyrene dihydrodiols in mouse skin and rat mammary gland. *Carcinogenesis*, 12: 1939–1944.
- [84] Toth B (1980). Tumorigenesis by benzo[a]pyrene administered intracolonicly. *Oncology*, 37: 77–82.
- [85] Anderson LM, Priest LJ, Deschner EE, Budinger JM (1983). Carcinogenic effects of intracolonic benzo[a]pyrene in  $\beta$ -naphthoflavone-induced mice. *Cancer Letters*, 20: 117–123.

- [86] Naslund I, Rubio CA, Auer GU (1987). Nuclear DNA changes during pathogenesis of squamous cell carcinoma of the cervix in 3,4-benzopyrene-treated mice. *Analytical and Quantitative Cytology*, 9: 411–418.
- [87] Rossi L, Barbieri O, Sanguineti M *et al.* (1983). Carcinogenic activity of benzo[a]pyrene and some of its synthetic derivatives by direct injection into the mouse fetus. *Carcinogenesis*, 4: 153–156.
- [88] Oguri T, Singh SV, Nemoto K, Lazo JS(2003). The carcinogen (7R,8S)-dihydroxy-(9S,10R)-epoxy-7,8,9,10-tetrahydrobenzo[a]pyrene induces Cdc25B expression in human bronchial and lung cancer cells. *Cancer Res*,63: 771–775.
- [89] Damiani LA, Yingling CM, Leng S *et al*(2008). Carcinogen-induced gene promoter hypermethylation is mediated by DNMT1 and causal for transformation of immortalized bronchial epithelial cells. *Cancer Res*, 68: 9005–9014.
- [90] Chen JH, Chou FP, Lin HH, Wang CJ(2005). Gaseous nitrogen oxide repressed benzo[a]pyrene-induced human lung fibroblast cell apoptosis via inhibiting JNK1 signals. *Arch Toxicol*, 79: 694–704.
- [91] Xiao H, Rawal M, Hahm ER, Singh SV(2007). Benzo[a] pyrene-7,8-diol-9,10-epoxide causes caspase-mediated apoptosis in H460 human lung cancer cell line. *Cell Cycle*, 6: 2826–2834.
- [92] Wojdani A & Alfred LJ (1984). Alterations in cell-mediated immune functions induced in mouse splenic lymphocytes by polycyclic aromatic hydrocarbons. *Cancer Res*, 44: 942–945.
- [93] Urso P & Gengozian N (1984). Subnormal expression of cell mediated and humoral immune responses in progeny disposed toward a high incidence of tumors after in utero exposure to benzo[a]pyrene. *J Toxicol Environ Health*, 14: 569–584.
- [94] Burgoon L, Cai C, Cooper G *et al*(2017). Toxicological Review of Benzo[a]pyrene. Integrated Risk Information System National Center for Environmental Assessment Office of Research and Development U.S. Environmental Protection Agency Washington, DC.1-234.
- [95] Einem Lindeman T, Poirier MC, Divi RL (2011). The resveratrol analogue, 2,3',4,5'-tetramethoxystilbene, does not inhibit cyp gene expression, enzyme activity and benzo[a]pyrene-DNA adduct formation in MCF-7 cells exposed to benzo[a]pyrene. *Mutagenesis* 26:629-635.
- [96] Benford D, Dinovi M, Setzer RW (2010). Application of the margin-of-exposure (moe) approach to substances in food that are genotoxic and carcinogenic e.g.: Benzo[a]pyrene and polycyclic aromatic hydrocarbons. *Food Chem Toxicol* 48 Suppl1:S42-48.
- [97] Adeleye Y, Andersen M, Clewell R, Davies M, Dent M, Edwards S, *et al* (2015). Implementing toxicity testing in the 21st century (tt21c): Making safety decisions using toxicity pathways, and progress in a prototype risk assessment. *Toxicology*. 5;332:102-11.